Claudin and ovarian cancer

Klaudin ve over kanseri

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Abstract

Claudins are a family of proteins and the most important component of the tight junction. They constitute a paracellular barrier that controls the flow of molecules in the intercellular space of an epithelium. Although it seems that claudin should be down regulated in cancer cell, some claudins are, in fact highly elevated in various human cancers, including ovarian cancer. Whereas the functional significance of claudin overexpression in ovarian carcinoma is unclear, these proteins are important for migration, invasion, and survival of ovarian cancer cells. They clearly represent a general pathway in tumorigenesis, are a novel marker for ovarian cancer and may become a target for therapy or diagnosis of this disease.

Key words: Tight junction, occludin, claudin, ovarian cancer, Clostridium perfringens enterotoxin

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Introduction

Claudins are 20-27 kDa transmembrane proteins spanning the cellular membrane 4 times, with the N-terminal end and the C-terminal end both located in the cytoplasm, and two extracellular loops which show the highest degree of conservation. The first extracellular loop consists on average of 53 amino acids and the second one, being slightly smaller, of 24 amino acids. The N-terminal end is usually very short (4-10 amino acids), the C-terminal end varies in length from 21 to 63 and is necessary for the localisation of these proteins in the tight junctions. It is suspected that the cytoeines of individual or separate claudins form disulfide bonds. All human claudins (with the exception of Claudin 12) have domains that let them bind to PDZ domains of scaffold proteins.

Work from several groups has now confirmed the over expression of these proteins in ovarian cancer. To find the subtypes based on a specific marker is a lucrative idea in diagnosis and specific managements including targeting and prognosis, such as ER, PR and HER neu in breast cancer, cyclin D and some translocations in leukemias. Ovarian cancer is a killer disease of unknown aetiology. It is detected late and has no good treatment. So, effort is concentrated on the use of claudin for diagnosis and target of therapy in ovarian cancer. It is worthwhile to study claudin in detail to know its significance in ovarian cancer.

Tight junction

Specialized intercellular junctions, known as desmosomes and terminal bars were recognized as local modifications of the surface of adjacent yet separate cells, rather than as intercellular bridges (1,2) with an effect on cell-to-cell adhesion and epithelial permeability.

In the nineties, mammalian intercellular junctions were described and were categorized into four types: adherens junctions (AJ), desmosomes (DS), gap junctions (GJ), and tight junctions (TJ) (3). The major integral membrane proteins in Aj are glycoprotein, cadherins. The desmosomal integral membrane proteins are called desmogleins and desmocollins, similar in amino acid sequence to cadherins, and they fall into the cadherin superfamily. The integral membrane protein in Gj is a dense aggregation of multimeric channels, each of which consists of six identical proteins named connexins. TJ is an element of the epithelial and endothelial junctional complex. It seals cells to create the primary barrier to the diffusion of solutes through the paracellular pathway. It also works as a boundary between the apical and basolateral plasma membrane domains to create the polarization of epithelial and endothelial cells. It remains controversial whether the particles in the strands of TJ are predominantly lipidic in nature. However, detergent stability of TJ strands as
visualized by negative staining and freeze fracture proves that these elements are not composed solely of lipids. Tight junctions, together with adherens junctions and desmosomes, form the apical junctional complex in epithelial and endothelial cellular sheets. Adherens junctions and desmosomes are responsible for the mechanical adhesion between adjacent cells, whereas tight junctions are essential for the tight sealing of the cellular sheets, thus controlling paracellular ion flux and therefore maintaining tissue homeostasis. Tight junctions also play a crucial role in the maintenance of cell polarity by forming a fence that prevents lateral diffusion of membrane proteins and lipids, thereby maintaining the differential composition of the apical and basolateral domains.

**Occludin**

In the late nineties, ZO-1, a tight junction-associated protein, was derived from chick liver. This protein was not extractable from plasma membranes without detergent, suggesting that it is an integral membrane protein. When its cDNA was cloned and sequenced a new 504-amino acid, 55.9 kDa polypeptide with a hydrophilicity plot very similar to that of connexin was found. A search of the data base identified no proteins with significant homology to this membrane protein. Furuse et al. (3) designated this integral membrane protein localizing at tight junctions as “occludin.”

**Claudin**

Identification of claudin was regarded as the Holy Grail in this field. Although successive studies emphasized the importance of occludin in the structure and functions of TJs, it gradually became clear that the molecular architecture of TJ strands is more complex than expected. Especially, the fact that the occludin-deficient visceral endoderm cells still bear a well-developed network of TJ strands indicated that membrane proteins or lipid molecules as yet unidentified may constitute TJ strands (4). Another two distinct peptide sequences of 211 and 230 amino acids of about 22-kD were obtained in a similar experiment on chick liver. Hydrophilicity analysis suggested that both bore four transmembrane domains (Fig.1), although they did not show any sequence similarity to occludin. Immunofluorescence and immuno-electron microscopy revealed that both proteins were targeted to and incorporated into the TJ strand itself. Furuse et al. (3) designated them as “claudin-1” and “claudin-2”, respectively (12). Gradually in humans, 23 members of the family have been described.

**Claudin Expression in Cancer**

Decreased tight junction protein expression in cancer is consistent with the generally accepted idea that tumorigenesis is accompanied by a disruption of tight junctions, a process that may play an important role in the loss of cohesion, invasiveness, and lack of differentiation observed in cancer cells. Down-regulation of both occludin (in gastrointestinal tumors) and claudins (in breast cancer, gastric cancer as well as in colon cancer) is noticed in cancer. Claudin-7 is down-regulated in invasive breast cancer (and in head and neck cancer). Likewise, expression of claudin-4 in pancreatic cancer cells reduces invasiveness of these cells. Phosphorylation of tight junction proteins, including claudins, may also disrupt tight junction function in cancer5. Claudins down regulated are listed in Table 1.

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**Table 1. Down regulated claudins in various cancers**

<table>
<thead>
<tr>
<th>Cancer</th>
<th>Claudin gene</th>
<th>Expression</th>
<th>References</th>
</tr>
</thead>
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<tr>
<td>Breast</td>
<td>CLDN1</td>
<td>Down</td>
<td>7</td>
</tr>
<tr>
<td></td>
<td>CLDN7</td>
<td>Down</td>
<td>8</td>
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<tr>
<td></td>
<td>CLDN1, CLDN3, CLDN4</td>
<td>Variable</td>
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<tr>
<td>Breast and Paget’s disease</td>
<td>CLDN2, CLDN3, CLDN4, CLDN5</td>
<td>Variable</td>
<td>10</td>
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<tr>
<td>Colon</td>
<td>CLDN1</td>
<td>Variable</td>
<td>11</td>
</tr>
<tr>
<td>Gastric</td>
<td>CLDN4</td>
<td>Down</td>
<td>12</td>
</tr>
</tbody>
</table>

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*Figure 1. Claudins with four transmembrane domains. The extracellular loops target for therapy. The C-terminal is PDZ binding domain.*
Paradox
Many claudins, such as claudin-3 and claudin-4, are typically up-regulated in many cancers, suggesting that these proteins may have a positive effect on tumorigenesis. Recent work has shown that, in ovarian cells, expression of claudin-3 and claudin-4 may lead to an increase in invasion, motility, and cell survival, all characteristics important for metastasis. This was observed in advanced ovarian cancer but not in ovarian cystadenomas. Therefore, the functions of claudins may be highly tissue specific and may depend on the exact molecular circuitry of the cell. In addition, claudin-3 and claudin-4 have also been reported to be expressed in other cancers, such as breast, prostate, and pancreatic cancers. The overwhelming majority of studies published thus far report an overexpression of many claudins in various cancers (Table 2).

Thus, recent reports suggest that we may lack a full vision of the functional complexity of claudins and their possible functional connection to a larger protein family. Database searching reveals a large number of proteins with structural similarities to claudins but whose functional similarities remain largely unexplored. There was a lack of consistent nomenclature. Before their naming in 1998, three of the orthodox claudins had already been cloned and given other names and were characterized by nonbarrier functions.

All are not barrier-forming claudins, neither is it known if they have involvement in apoptosis and proliferation. Several orthodox claudins are not restricted to tight junctions; claudin-7, for example, is on the basolateral surface of cells in the kidney tubule epithelium. Many other claudins have large pools of protein on the lateral surface distinct from the barrier-forming TJ strands. The role and organization of extrajunctional claudin remain unclear. Even more enigmatic is the inclusion in the pfam0082 family of some subunits of voltage dependent calcium channels. Cytokines such as tumor necrosis factor alpha (TNFα) and interferon gamma (IFNγ) can modulate claudin. Moreover, claudins play a significant role in some autoimmune and hereditary diseases such as inflammatory bowel disease, hereditary deafness, multiple sclerosis, familial hypomagnesemia with hypercalciuria and nephrocalcinosis (FHHNC), diabetic retinopathy etc (31). It seems that not only tissue specificity but inflammatory and immunological conditions prevail, and variation of the functions of claudins may be responsible for a contradictory expression of claudins in many cancer.

Claudin and Ovarian Cancer
To date, research on claudin in ovarian cancer has been both on a basic and clinical level. Basic research involving cell line or animal experiments include a) detection of expression of many claudins b) exploring the mechanism of gene expression, methylation status, epigenetic study, influence of gonadotrophin and metabolic pathways like phospholipidation etc. In clinical research this effort is mainly limited to observation of tissue expressions of claudin in snap frozen or archival specimen for diagnosis and targeting by Clostridium perfringens enterotoxin, lipidoid siRNA etc.

Table 2. Up regulated claudins in various cancers

<table>
<thead>
<tr>
<th>Cancer</th>
<th>Claudin gene</th>
<th>Expression</th>
<th>References</th>
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</thead>
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<tr>
<td>Breast</td>
<td>CLDN3</td>
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<td>13</td>
</tr>
<tr>
<td></td>
<td>CLDN4</td>
<td>Up</td>
<td>13</td>
</tr>
<tr>
<td>Hepatocellular carcinoma</td>
<td>CLDN10</td>
<td>Up</td>
<td>14</td>
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<tr>
<td>Squamous cell carcinoma</td>
<td>CLDN1</td>
<td>Up</td>
<td>15</td>
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<tr>
<td></td>
<td>CLDN4</td>
<td>Up</td>
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<td>CLDN3</td>
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<tr>
<td></td>
<td>CLDN4</td>
<td>Up</td>
<td>16,18,19</td>
</tr>
<tr>
<td></td>
<td>CLDN16</td>
<td>Up</td>
<td>20</td>
</tr>
<tr>
<td>Pancreatic</td>
<td>CLDN4</td>
<td>Up</td>
<td>21,22,23,24</td>
</tr>
<tr>
<td>Pancreatic (intraductal papillary mucinous neoplasms)</td>
<td>CLDN4</td>
<td>Up</td>
<td>25</td>
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<tr>
<td></td>
<td>CLDN18.2</td>
<td>Up</td>
<td></td>
</tr>
<tr>
<td>Prostate</td>
<td>CLDN3</td>
<td>Up</td>
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</tr>
<tr>
<td></td>
<td>CLDN4</td>
<td>Up</td>
<td>26</td>
</tr>
<tr>
<td>Thyroid papillary cancer</td>
<td>CLDN10</td>
<td>Up</td>
<td>27</td>
</tr>
<tr>
<td>Endometrial Cancer</td>
<td>CLDN4</td>
<td>Up</td>
<td>28</td>
</tr>
<tr>
<td>Oesophageal tumour</td>
<td>CLDN18.2</td>
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<td>29</td>
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<td>non-small cell lung cancer: Squamous cell carcinoma</td>
<td>CLDN1</td>
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<td>Adenocarcinoma</td>
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<tr>
<td></td>
<td>CLDN5</td>
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1. Important basic researches

Serial analysis of gene expression (SAGE) was used to generate global gene expression profiles from various ovarian cell lines and tissues, including primary cancers and ovarian surface epithelia cells, in archival material and cystadenoma cells (16). This showed upregulation of claudin-3 and claudin-4 gene and this was further validated through immunohistochemical analysis. It was only in the previous year that SAGE had detected the tag for the *Claudin-7* gene 12 times in the tumor cell lines for the first time, whereas no *Claudin-7* tags were found in the normal breast cultures (32). The earlier group tried to account for the significance of its upregulation. Although *CLDN4* overexpression is well established, the mechanisms responsible for this abnormal regulation remain unknown. In a study, they delineated a small region of the *CLDN4* promoter critical for its expression. This region contains two Sp1 sites, both of which are required for promoter activity. However, because of the ubiquitous expression of Sp1, these sites, although necessary, are not sufficient to explain the patterns of *CLDN4* gene expression in various ovarian tissues. *CLDN4* promoter is further controlled by epigenetic modifications of the Sp1-containing critical promoter region. Cells that overexpress *CLDN4* exhibit low DNA methylation and high histone H3 acetylation of the critical *CLDN4* promoter region, and the reverse is observed in cells that do not express *CLDN4*. Moreover, the *CLDN4*-negative cells can be induced to express *CLDN4* through treatment with demethylating and/or acetylating agents. Because *CLDN4* is elevated in a large fraction of ovarian cancer, they opined that the mechanism leading to deregulation may represent a general pathway in ovarian tumorigenesis and may lead to novel strategies for therapy and an overall better understanding of the biology of this disease (33).

In a recent phosphorylation study, analyses using PKC inhibitors, siRNA and immunofluorescence have shown that PKC-epsilon, an isoform typically expressed in ovarian cancer cells, may be important in the TPA-mediated claudin-4 phosphorylation and weakening of the TJs (34). In *CLDN3* promoter it was possible to identify a minimal region containing a Sp1 site crucial for its activity (35). In addition, this group found that the *CLDN3* promoter is regulated through epigenetic processes as in *CLDN4*. Interestingly, in vitro binding experiments, as well as chip assays show that Sp1 binds the unmethylated promoter much more efficiently, providing a mechanism for *CLDN3* silencing in non-expressing cells. siRNA-mediated knockdown of Sp1 led to a significant decrease of *CLDN3* expression at both the mRNA and protein levels, demonstrating a crucial role for this transcription factor in the regulation of *CLDN3* (36). Animal experiments from Canada showed that, in the absence of FSH-R signaling, claudin-3, claudin-4, and claudin-11 are selectively upregulated, whereas claudin-1 decreases in ovarian surface epithelium. In vitro experiments using a mouse ovarian surface epithelial cell line derived from wild-type females reveal a direct hormonal influence on claudin proteins. Although recent studies suggest that cell junction proteins are differentially expressed in ovarian tumors in women, the etiology of such changes remains unclear. They suggest an altered hormonal environment resulting from FSH-R loss as a cause of early changes in tight junction proteins that predispose the ovary to late-onset tumors that occur with aging (36).

2. Clinical research

a) Diagnosing ovarian cancer, tissue marker

The result of the SAGE study on ovarian cancer was established in a detailed analysis of *CLDN3* and *CLDN4* expression in a panel of ovarian tumors of various subtypes and cell lines. RNA was obtained from a panel of 39 microdissected epithelial ovarian tumors of various histological subtypes for real-time reverse transcription-PCR analysis. In addition, a total of 70 cases of ovarian carcinomas, ovarian cysts, and normal ovarian epithelium from a tissue array were analyzed and validated by immunohistochemistry (21).

Affymetrix human genome arrays (U95 series) was utilized to analyze differences in gene expression of 41,441 known genes and expressed sequence tags between five pools of normal ovarian surface epithelial cells (OSE) and 42 epithelial ovarian cancers of different stages, grades, and histotypes (17). The 3-fold up-regulated genes were analyzed using recursive descent plot analysis (RDPA), and the combination of elevated claudin 3 gene (*CLDN3*) and elevated VEGF distinguished the cancers from normal OSE. A combination of *CLDN3*, *CA125*, and *MUC1* (mucin-1 transmembrane) stained 99.4% of 158 cancers, and all of the tumors were detected with a combination of *CLDN3*, *CA125*, *MUC1*, and VEGF. Thus, a limited number of markers in combination might identify >99% of epithelial ovarian cancers despite the heterogeneity of the disease.

Total RNA was extracted and transcription profiling performed in another experiment (37) using the Eos Hu03, a customized Affymetrix GeneChip oligonucleotide microarray (Affymetrix, Santa Clara, CA). The expression patterns of three integral membrane proteins, discoidin domain receptor 1 (DDR1), claudin 3 (*CLDN3*), and epithelial cell adhesion molecule, all of which are involved in cell adhesion, were evaluated in a cohort of 158 primary EOC using immunohistochemistry identifying DDR1 and *CLDN3* as new biomarkers of EOC. Two other DNA microarray affymetrix study results corroborated claudin 3 and 4 gene overexpression in ovarian cancer. They used Hierarchical clustering of the expression data and Youden’s misclassification index for each gene marker via pairwise tissue comparisons respectively (18, 19).

No new claudins were implicated with ovarian cancer in 2005. The Professor Bast Jr. group, in search of potential markers that complement expression of *CA125* in epithelial ovarian cancer, included claudin 3 with at least ten other markers at the level of tissue expression (38).

In 2006 it was substantiated by both Swedish and Italian groups that both claudin 3 and 4, even though they differ in expression during ovarian malignant transformation, might be used as novel markers for ovarian tumours (39,40). In another experiment, strong expression of claudins 1, 4, and 7 was seen in benign and malignant epithelial ovarian tumors. Expression of claudin 5, reported to be mainly present in endothelial cells, was seen in ovarian epithelial tumors, but with a significantly lower frequency than claudins 1, 4, and 7. On the contrary, sex-cord stromal tumors and cysts, such as fibromas/thecomas, Sertoli-Leydig cell tumors, granulosa cell tumors, and follicular and luteinized cysts were mainly negative for claudins 1, 4, 5, and 7 (41). Ovarian/primary peritoneal serous carcinoma (OC/ PPC) and diffuse peritoneal malignant mesothelioma (DMPM) are highly aggressive tumors. Claudins 3, 4, and 6 overexpression...
was noticed in them (42). Claudin 4 was among few proteins whose expression profiles correlated with cisplatin resistance in ovarian cancer cells. Several proteins may be involved in modulating response to cisplatin and have a potential as markers of treatment response or treatment targets (43). Expression of claudin-3 or claudin-7 is specific for adenocarcinoma and rules out the diagnosis of cells as mesothelial and the absence of claudin-1 expression excludes ovarian carcinoma as the possible origin of metastatic adenocarcinoma. Claudins may, therefore, be of diagnostic value in biopsy and effusion cytology (44). Kaplan-Meier survival analysis showed that serous ovarian adenocarcinoma patients with high CLDN3 expression had a substantially shorter survival (P=0.027). Multivariate analysis demonstrated that CLDN3 overexpression is an independent negative prognostic factor (45). Expression for all cell-junction proteins with a typical honeycomb-staining pattern in the serous adenocarcinomas and not Clear-cell and endometrioid adenocarcinomas prove that Serous adenocarcinomas form functional TJs in vitro (46).

CLDN-7 transcript was found significantly overexpressed in both primary and metastatic EOCs compared to normal human ovarian surface epithelium cell lines (fold change=111.4, P<0.001) by reverse transcription-polymerase chain reaction, regardless of the histologic type, the grade of differentiation, and the pathologic stage of the disease (47). Moreover, a strong immunoreactivity for CLDN-7 was detected in EOC cells present in ascites fluids, whereas ascites-derived inflammatory cells, histiocytes, and reactive mesothelial cells were negative. With the exception of claudin-4, claudins are upregulated in OC effusions compared with solid tumors, in agreement with previous data for cadherins and integrins in this cancer type, suggesting a prosurvival role for these surface molecules. Claudin-3 and claudin-7 expression in effusions independently predicts poor survival in OC (48). Increased expression of claudin-3 and claudin-4 may contribute to the aggressive phenotype of endometrial cancer of serous papillary or clear-cell histology also and suggests their potential utility as diagnostic biomarkers and possible targets for therapeutic intervention (28).

b) Targeting claudin for therapy

**Clostridium perfringens enterotoxin receptor**

The peptide toxin Clostridium perfringens enterotoxin (CPE) has been shown to bind to claudin-3 and -4, but not to claudin-1 or -2 (49). Claudin-1/claudin-3 chimeric molecules showed that the second extracellular loop of claudin-3 conferred CPE sensitivity on L fibroblasts. The second extracellular loop is the site through which claudin-3 interacts with CPE on the cell surface. Claudins 3 and 4 have been described as the low- and high-affinity receptors, respectively, for the cytocytic Clostridium perfringens enterotoxin (CPE) (50). Their sensitivity to CPE treatment was seen in vitro when 100% (17 of 17) of the primary ovarian tumors tested overexpressed one or both CPE receptors by quantitative reverse transcription-PCR. All ovarian tumors showed a dose-dependent cytotoxic effect to CPE in vitro. All primary ovarian tumors tested died within 24 hours of exposure to 3.3 microg/mL CPE in vitro. CPE therapy in SCID mouse xenografts in a highly relevant clinical model of chemotheraphy-resistant freshly explanted human ovarian cancer (i.e., OVA-1) showed that multiple i.p. administration of sublethal doses of CPE every 3 days significantly inhibited tumor growth in 100% of mice harboring 1 week established OVA-1. Claudin-4 overexpression in epithelial ovarian cancer (EOC) does not correlate with survival or other clinical endpoints and was found to be associated with hypomethylation. Claudin-4 overexpression correlates with Rb and C-CPE. Treatment of EOC cells with C-CPE significantly decreased Rb in a dose- and claudin-4-dependent noncytotoxic manner. Thus, C-CPE treatment of EOC cells may lead to altered TJ function induced cytotoxicity, (51) and hence is a therapeutic measure.

**Pan-cancer target**

CLDN18.2 is retained on malignant transformation and is expressed in a significant proportion of primary gastric cancers and the metastases thereof. In addition, its orthotopic expression was found in pancreatic, esophageal, ovarian, and lung tumors, correlating with distinct histologic subtypes. The activation of CLDN18.2 depends on the binding of the transcription factor cyclic AMP-responsive element binding protein to its unmethylated consensus site. Most importantly, it was possible to raise monoclonal antibodies that bind to CLDN18.2. Its highly restricted expression pattern in normal tissues, its frequent ectopic activation in a diversity of human cancers, and the ability to specifically target this molecule at the cell surface of tumor cells, qualify CLDN18.2 as a novel, highly attractive pan-cancer target for the antibody therapy of epithelial tumors including ovarian cancer (52).

**Claudin-3 gene silencing with siRNA**

In a recent US study (53), intratumoral injection of lipidoid/CLDN3 siRNA into OVCAR-3 xenografts resulted in dramatic silencing of CLDN3, significant reduction in cell proliferation, reduction in tumor growth, and a significant increase in the number of apoptotic cells. Intrapitoneal injection of lipidoid-formulated CLDN3 siRNA resulted in a substantial reduction in tumor burden in Misis/Tg transgenic mice and mice bearing tumors derived from mouse ovarian surface epithelial cells. It has been reported that CLDN3 is expressed at very low levels in several normal tissues in humans including the lungs, kidneys, breast, uterus, pancreas, and thyroid. Colon, small bowel, and prostate are the only normal tissues that show appreciable expression. An I.p. administration of CLDN3 siRNA formulations may reduce the concern of adverse effects of silencing CLDN3 in healthy tissues that reside outside the peritoneum. Lipidoid-formulated CLDN3 siRNA has potential as a therapeutic agent for ovarian cancer.

**Disruption of TJs**

The C terminus of claudin-3 was seen to be an excellent substrate for cAMP-dependent protein kinase (PKA) at threonine 192. Overexpression of the protein containing a T192D mutation, mimicking the phosphorylated state, resulted in a decrease in TJ strength in ovarian cancer cell line Ovca433. This may provide a mechanism for the disruption of TJs in this cancer causing cytotoxicity (54). Another report showed that the claudin-expressing ovarian epithelial cells were found to have increased matrix metalloproteinase-2 (MMP-2) activity indicating that claudin-mediated increased invasion might be mediated through the activation of MMP proteins. However,
siRNA inactivation of claudins in ovarian cancer cell lines did not have a significant effect on the high endogenous MMP-2 activity present in these cells, showing that malignant cells have alternative or additional pathways to fully activate MMP-2 (55).

Claudin crosstalk
Gene expression mediated by the promoter of claudin-2 may be regulated by factors involved in Wnt signaling (56). Moreover, a functional crosstalk between Wnt signaling and transcriptional activation related to caudal-related homeobox (Cdx) proteins could be demonstrated in the regulation of claudin-2 promoter-mediated gene expression.

Although formed by different molecules, tight junctions (TJs) and adherens junctions (AJs) are functionally and structurally linked, but the signalling pathways behind this interaction are unknown. A cell-specific mechanism of crosstalk between these two types of structure was shown when endothelial VE-cadherin at AJs upregulated the gene encoding the TJ adhesive protein claudin-5 (57). This effect required the release of the inhibitory activity of forkhead box factor FoxO1 and the Tcf-4-beta-catenin transcriptional repressor complex. Vascular endothelial (VE)-cadherin acts by inducing the phosphorylation of FoxO1 through Akt activation and by limiting the translocation of beta-catenin to the nucleus. These results offer a molecular basis for the link between AJs and TJs and explain why VE-cadherin inhibition may cause a marked increase in permeability. These findings might have bearing in ovarian cancer.

Conclusion
Unusual expression patterns of claudins suggest an utility for detection, diagnosis, and treatment of drug-resistant cancers. Hopefully, new experiments will pave the way for proper utilization of knowledge already gathered to the benefit of advancement in management of many diseases, including ovarian cancer. However, clinical trials will be required to establish these potential. Until then, the interesting nature of the subject, and need for basic and clinical research on claudins cannot be overemphasized as claudin is likely to remain valuable for providing important insights into normal and neoplastic cellular physiology. The paradox of the findings has to be solved, and diagnosis and targeting has to be established in future.

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Conflict of interest
None declared

References


