Introduction

Important alterations in maternal metabolism and increased insulin resistance coincide with the progressive accumulation of adiposity during the course of normal pregnancy. Insulin resistance shows an increase in the late second trimester to levels that are observed in type 2 diabetes mellitus (T2DM) (1). Most pregnant women remain normoglycemic due to adequate beta-cell compensation for this higher insulin secretion. When the beta-cell compensation for insulin resistance and hepatic glucose production is inadequate, gestational diabetes mellitus (GDM) ultimately develops (1). Moreover, 10% to 50% of GDM cases are reported to develop T2DM in the postpartum period (2).

Gestational diabetes mellitus is defined as a condition of carbohydrate intolerance with onset or first recognition during pregnancy (3). The American Diabetes Association states that the incidence of GDM is between 1% and 14%, and it complicates almost 7% of pregnancies (4). GDM carries numerous risks for mothers, fetuses, and even offspring. GDM causes vascular and obstetric complications, including diabetic nephropathy, retinopathy, macrosomia, increased operative deliveries, and unexplained fetal demise. Neonatal complications, such as hypoglycemia, hypocalcemia, jaundice, respiratory distress syndrome, and cardiomyopathy, are also more prevalent. Offspring born to women with diabetes have a 1% to 3% risk of cardiovascular and metabolic disorders (2, 5).

Maternal adiposity is an important, modifiable risk factor for the development of GDM (6). Adipose tissue is not only involved in energy storage, but also functions as an active endocrine organ (7). Recent evidence supports the crucial roles of specific hormones and adipokines (i.e., cytokines) secreted by adipose tissue. Among those identified to date are adiponectin, leptin, resistin, tumor necrosis factor alpha (TNF-alpha), progranulin, omentin, chemerin, retinol binding protein-4 (RBP-4), and visfatin.

Although adipokine chemerin has conventionally been associated with regulation of adaptive and innate immunity, it also induces insulin resistance and impairs glucose tolerance. Chemerin is a novel adipokine that is secreted by various tissues, especially white adipose tissue; it regulates insulin sensitivity in adipocytes and skeletal muscle (8, 9). Increased
levels of chemerin are detected in obesity and are associated with multiple components of metabolic syndromes, including body mass index (BMI), triglycerides, high density cholesterol, hypertension, inflammation, and markers of liver pathology (10). Chemerin is also expressed in human placenta (11). Serum levels of chemerin correlate significantly with systemic markers of inflammation, such as TNF-alpha, interleukin 6, and C-reactive protein (12). Overall, these findings suggest that chemerin levels are related to adiposity and glucose metabolism. They also represent a possible link between obesity and the development of GDM.

Retinol binding protein-4 is a blood carrier protein for retinol that is synthesized in hepatocytes and adipocytes (13). Increased levels of RBP-4 have been demonstrated in several metabolic conditions, including obesity, insulin resistance, polycystic ovary syndrome, and cardiovascular disease (14). Also, RBP-4 is believed to induce expression of enzymes involved in gluconeogenesis in hepatocytes and to impair insulin signaling pathways in skeletal muscle (15).

Visfatin is predominantly secreted by visceral tissue; however, it is also found in skeletal muscle, liver, bone marrow, lymphocytes, and placenta (16). Visfatin promotes adipogenesis and exerts insulin-mimetic effects (17). It also upregulates production of proinflammatory cytokines by monocytes (18). Circulating levels of visfatin are increased in patients with type 1 and 2 DM and obesity (13, 19). However, the association of visfatin with GDM is still unclear (20).

Only a few adipokines have been investigated with respect to their involvement in GDM. Evidence in the existing literature does not support clear roles of chemerin, RBP-4, and visfatin in the prediction of GDM. In this study, we aimed to investigate the association of maternal serum levels of chemerin, RBP-4, visfatin, and insulin with GDM. Therefore, we performed a prospective cross-sectional study in pregnant women with GDM and with normal glucose tolerance.

Material and Methods

Study population
This cross-sectional study was conducted in the Obstetrics and Gynecology Department of Hitit University between March 2015 and September 2015. The ethics committee of Ankara Numune Hospital approved the study, which was in accordance with the Declaration of Helsinki, 2013 Brazil version (20796219-724.131). All participants gave written informed consent for the study. The patients were 18 to 35 years of age and had singleton pregnancies. Pregnant women who presented themselves to our obstetrics department were screened between 24 and 28 weeks of gestation for GDM according to the recommendations of the American College of Obstetricians and Gynecologists (ACOG) (24). Briefly, all pregnant women in the low risk group were evaluated with a 50-g glucose challenge test (GCT). Women with serum glucose ≥140 mg/dL at 1 h after GCT were subjected to a 100-g oral glucose tolerance test (OGTT). Serum glucose concentrations were measured at 0, 1, 2, and 3 h after glucose ingestion. The diagnosis of GDM was based on the criteria of Carpenter and Couston, in which, after a 100-g oral glucose load, 2 or more of the following plasma values must be obtained: fasting ≥95 mg/dL, 1h ≥180 mg/dL, 2h ≥155 mg/dL, and 3h ≥140 mg/dL (21). The estimation of pregnancy duration was based on routine ultrasonographic examination performed in the first trimester. BMI was calculated using pregnancy weight and height, which were recorded at the time of blood sampling. The exclusion criteria were as follows: (1) smoking, (2) a history of diabetes mellitus and/or GDM, (3) a history of chronic disease, (4) a history of congenital malformation, (5) a family history of DM. A total of 158 pregnant women met the inclusion criteria and were divided into two groups: 76 were in the GDM group, and 82 were in the control group. The demographic characteristics and biochemical parameters of the study population, including age, BMI, and gestational age, were recorded in the second trimester.

Assays
Blood samples for adipokines and insulin were obtained from the antecubital vein after overnight fasting between 8:00 A.M. and 10:00 A.M. The samples for adipokines and insulin were centrifuged (1500 g for 25 min), and the serum was immediately stored at -80 °C until analysis. Serum glucose was determined daily using the glucose hexokinase method (Siemens Healthcare Diagnostic Limited; Camberley, UK). The serum chemerin concentration was measured by the enzyme-linked immunosorbent assay (ELISA) method (Biovendor, Biovendor-Laboratori Mediicina; Brno, Czech Republic). Serum RBP-4 concentration was determined by ELISA (Immundiagnostik, Immundiagnostik AG; Bensheim, Germany). Serum visfatin level was also determined by ELISA (Cusabio, Cusabio Biotech Co. Ltd.; Hubei, China). Serum insulin concentration was measured by chemiluminescence assay (Advia Centaur, Siemens Medical Solutions Diagnostics; Tarrytown, USA). Homeostasis model assessment-insulin resistance (HOMA-IR) was calculated using the following formula: Plasma glucose (mg/dL)×fasting plasma insulin (IU/mL) in the fasting state divided by 405 (22).

Statistical analysis
Data were analyzed using SPSS (Statistical Package for the Social Sciences) software version 21 (SPSS Inc.; Chicago, IL, USA). Continuous variables were first evaluated for normality of statistical distribution by the Shapiro-Wilk test. As the continuous variables were not normally distributed, a non-parametric method (Mann-Whitney U test) was used to perform the statistical analysis. Descriptive statistics were expressed as the median (minimum-maximum) and number (percentage %). Spearman correlation tests were used to determine the correlations of continuous variables. P values <0.05 were considered to be significant in all comparisons.

Results
The maternal demographic characteristics and biochemical parameters of the study participants are summarized in Table 1; all continuous variables are given as the median (minimum-maximum). There were no differences in age and gestational
Table 1. Comparison of maternal demographic characteristics and biochemical parameters in the gestational diabetes mellitus (GDM) and control groups

<table>
<thead>
<tr>
<th></th>
<th>GDM group</th>
<th>Control group</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td>Number</td>
<td>76 (48.1%)</td>
<td>82 (51.9%)</td>
<td></td>
</tr>
<tr>
<td>Maternal age (years)</td>
<td>29 (19-35)</td>
<td>26 (18-35)</td>
<td>0.058</td>
</tr>
<tr>
<td>Gestational age (weeks)</td>
<td>26.5 (24-28)</td>
<td>25 (24-28)</td>
<td>0.820</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>33.25 (22.80-52.20)</td>
<td>26.43 (19.10-47.00)</td>
<td>0.000*</td>
</tr>
<tr>
<td>Chemerin (ng/mL)</td>
<td>3.64 (2.37-7.43)</td>
<td>3.47 (2.19-4.63)</td>
<td>0.100</td>
</tr>
<tr>
<td>RBP-4 (mg/mL)</td>
<td>15.29 (10.08-19.63)</td>
<td>14.91 (10.64-31.16)</td>
<td>0.871</td>
</tr>
<tr>
<td>Visfatin (ng/mL)</td>
<td>0.07 (0.03-0.44)</td>
<td>0.07 (0.03-0.47)</td>
<td>0.886</td>
</tr>
<tr>
<td>Insulin (mIU/mL)</td>
<td>14.94 (1.39-32.26)</td>
<td>9.87 (3.53-23.98)</td>
<td>0.000*</td>
</tr>
<tr>
<td>HOMA-IR</td>
<td>3.73 (0.33-8.40)</td>
<td>1.77 (0.60-4.59)</td>
<td>0.000*</td>
</tr>
</tbody>
</table>

BMI: body mass index; RBP-4: retinol binding protein-4; HOMA-IR: homeostasis model assessment of insulin resistance
*p values indicate statistical significance (p<0.05).
Values are shown as median (minimum-maximum).

Table 2. Correlation analyses of data

<table>
<thead>
<tr>
<th></th>
<th>Chemerin</th>
<th>Visfatin</th>
<th>Insulin</th>
<th>HOMA-IR</th>
</tr>
</thead>
<tbody>
<tr>
<td>RBP-4</td>
<td>r 0.251</td>
<td>-0.071</td>
<td>-0.018</td>
<td>0.021</td>
</tr>
<tr>
<td></td>
<td>p 0.026*</td>
<td>0.534</td>
<td>0.873</td>
<td>0.854</td>
</tr>
<tr>
<td>Chemerin</td>
<td>r 0.192</td>
<td>0.196</td>
<td>0.161</td>
<td></td>
</tr>
<tr>
<td></td>
<td>p 0.090</td>
<td>0.083</td>
<td>0.155</td>
<td></td>
</tr>
<tr>
<td>Visfatin</td>
<td>r 0.143</td>
<td>0.071</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>p 0.207</td>
<td>0.535</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Insulin</td>
<td>r 0.868</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>p 0.000*</td>
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</table>

RBP-4: retinol binding protein-4; HOMA-IR: homeostasis model assessment of insulin resistance
*p values indicate statistical significance (p<0.05).

Discussion

Various adipokines contribute to diabetogenic resistance to insulin, especially during the last half of pregnancy. This study mainly focused on potential alterations of specific adipokine concentrations (chemerin, RBP-4, and visfatin) in pregnant women with GDM. Here, we demonstrate that there were no significant differences in these adipokines between pregnant women with GDM and healthy pregnant women in the second trimester. However, the women with GDM were more likely to be overweight compared to matched healthy controls. Chemerin has been proposed to be an insulin-sensitizing adipokine; its secretion has been demonstrated to increase, presumably as a compensatory mechanism, in insulin-resistant subjects (11). However, our results demonstrate the exact opposite. Various studies have evaluated the correlation of chemerin with clinical parameters in pregnant women. To date, no study has precisely determined the predictive value of chemerin concentration for risk of GDM. A recent study from Germany reported that circulating levels of chemerin did not show a significant difference between pregnant women with GDM and healthy pregnant controls (1). Garces et al. (23) found no significant association of insulin and HOMA levels with chemerin, as did other independent studies (11, 24, 25). We found similar results for chemerin in pregnant women with GDM; however, other studies presented an association of chemerin with insulin resistance in pregnancy (26, 27). Overall, it appears that chemerin may not be useful in predicting GDM. As previously outlined, inconsistent data has been published concerning the association of RBP-4 with GDM. Some, but not all, studies have proposed that RBP-4 is positively and independently correlated with insulin resistance (28). Many studies demonstrated increased RBP-4 concentrations in GDM (14, 29), while others did not show a relationship of chemerin with GDM or normal glucose tolerance, as in our study (30, 31). One study even reported that RBP-4 concentration was decreased in GDM (26). A study was conducted by Wójcik et al. (13) concerning the relationship between adipose tissue hormones and GDM. This findings of this study conflicted with those in previous reports.

In the present study, we also found that the visfatin levels of pregnant women with and without GDM were not significantly different. Current evidence regarding the association of visfatin concentration with GDM is contradictory. Some studies demonstrated increased serum levels of visfatin in women with GDM (11, 32, 33), while others reported that visfatin concentrations were significantly lower in women with GDM (34, 35). Contrary to our study, Lewandowski et al. (32) and Akturk et al. (35) reported a significant correlation between visfatin and HOMA. It is presumed that this discrepancy may be related to differences in gestational duration at the time of sampling, the variety of diagnostic criteria for GDM, or even racial differences (36). Furthermore, Morgan et al. (37) suggested that visfatin may act locally as a paracrine/autocrine agent and not as a hormone. However, the exact mechanism remains unclear. In a study by Rezvan, no correlation of visfatin was observed with insulin or HOMA (38).
Despite the high number of studies evaluating the role of adipokines in GDM in the existing literature, interpretation of the results is somewhat laborious for several reasons. First, the diagnostic criteria for GDM vary greatly. Second, the gestational age at the study times ranges from early first trimester to late third trimester. Third, diverse assay methods may also cause heterogenous results. Obesity is accompanied by altered secretion of adipokines from adipose tissue (39, 40). Adipokine levels are usually higher in obese women. Although we demonstrated that pregnant women with GDM had higher BMI values than healthy pregnant women, there were no differences in the levels of the studied adipokines between the two groups. This may be due to the regulation of various adipokines by pregnancy or to insufficient matching of control and GDM patients for BMI.

Limitations in the present study that should be mentioned are the relatively small number of participants in each group and the assessment of maternal adipokines only in the second trimester. Furthermore, we did not analyze adipokines during the pre-pregnancy period, during the course of pregnancy, and in the postpartum period. Finally, power analysis was not performed due to the limited financial resources of this study, which was supported by the Scientific Research Unit of Hitit University.

In conclusion, despite these limitations, our study supports that serum chemerin, RBP-4, and visfatin levels in pregnant women with GDM do not differ from those of healthy pregnant women. We suggest that long-term observations of adipokines during the pre-pregnancy, pregnancy, and postpartum periods would increase our understanding of the pathogenesis of GDM. Therefore, further prospective studies are essential to elucidate the contribution of adipokines to GDM and the positive correlation between maternal RBP-4 and chemerin.

**Ethics Committee Approval:** Ethics committee approval was received for this study from the ethics committee of Ankara Numune Hospital (20796219-724.131).

**Informed Consent:** Written informed consent was obtained from patients who participated in this study.

**Peer-review:** Externally peer-reviewed.

**Author Contributions:** Concept - Ü.G.; Design - Ü.G.; Supervision - T.G.; Data Collection and/or Processing - Ü.G., F.K.K.; Analysis and/or Interpretation - Ü.G., C.T.; Literature Search - Ü.G., C.T.; Writing Manuscript - Ü.G.; Other - T.G.

**Acknowledgements:** The authors would like to thank the staff at Hitit University for collecting the data and their assistance.

**Conflict of Interest:** No conflict of interest was declared by the authors.

**Financial Disclosure:** The authors declared that this study has received financial support from the Scientific Research Unit of Hitit University, Çorum, Turkey.

**References**