Case report

A Novel Homozygous Mutation of the Acid-Labile Subunit (IGFALS) Gene in a Male Adolescent

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What is already known?
1. Patients with ALS (IGFALS) mutations have markedly decreased IGF-1, and extremely low IGFBP-3 levels
2. Although patients with ALS deficiency show moderate short stature, the phenotype of ALS deficiency is quite variable. Microcephaly, delay in puberty, insulin resistance, and reduced bone mineral density (BMD) have been shown in some patients.

What this study adds?
1. Novel homozygous frameshift mutation in IGFALS (p.Ser555Thrfs.19) causes short stature and delayed puberty but ultimately, with an obvious pubertal growth acceleration and good pubertal height gain, ending up in a normal adult height comparable to the target height.
2. Heterozygous carriers of this mutation have normal prenatal growth, puberty, insulin metabolism and BMD.

ABSTRACT
Acid-labile subunit (ALS) forms ternary complexes with insulin like growth factor-1 (IGF-1) and IGF-binding protein-3 (IGFBP-3) and is essential for normal circulating IGF-1 levels. The IGFALS gene encodes the ALS, and mutations in IGFALS cause ALS deficiency. We describe a patient with ALS deficiency with a novel homozygous frameshift mutation in IGFALS presenting with short stature and delayed puberty but ultimately achieving an adult height (AH) comparable to his target height (TH). A 15 3/12 year old boy presented with short stature (149.9 cm, –3.04 SDS). The patient had a low circulating IGF-1 level, extremely low IGFBP-3 level, insulin resistance and osteopenia. The peak growth hormone (GH) response to GH stimulation test was high (31.6 ng/mL). Sequencing of IGFALS revealed a novel homozygous frameshift mutation (p.Ser555Thrfs.19). His mother and elder sister were heterozygous carriers. Although he had delayed puberty and short stature at the onset of puberty, he reached his TH and an AH similar to those of his heterozygous mother and sister. The heterozygous carriers had normal or low IGF-1 levels and low IGFBP-3 levels but not as extremely low as that of the patient. They had normally timed puberty, insulin metabolism and bone mineral density (BMD). The phenotype of ALS deficiency is quite variable. Despite short stature and delayed puberty, patients can end up with normal pubertal growth and AH. ALS deficiency may cause osteopenia and hyperinsulinemia. Heterozygous carriers may have normal prenatal growth, puberty, insulin metabolism and BMD.

Keywords: Short stature, Acid-labile subunit deficiency, IGFALS gene mutation, Primary IGF-1 deficiency

Introduction
The majority of circulating insulin like growth factor-1 (IGF-1) is bound to IGF-binding proteins (IGFBP), mainly IGFBP-3, and the acid-labile subunit (ALS). ALS has a major role in stabilizing the 150-kDa ternary complex. The ternary complex extends the half-life of IGF-1 from 10 minutes in the free form to more than 12 hours (1,2). Therefore, ALS is necessary to maintain normal circulating IGF-1 and IGFBP-3 levels. Patients with ALS (IGFALS) mutations have markedly decreased IGF-1, and extremely low IGFBP-3 levels, associated with normal or compensatory elevated growth hormone (GH) levels (3).
Patients typically show moderate short stature in contrast to other, more severe causes of primary IGF-1 deficiency. In addition to short stature, some other features have been reported in the phenotype of ALS deficiency. Although some of the clinical and laboratory features of patients remain controversial, microcephaly, delay in puberty, insulin resistance, and reduced bone mineral density (BMD) have been shown in some patients (4-15).

IGFALS is located on chromosome 16p13.3 and encodes the 85-kDa ALS glycoprotein. ALS is produced by the liver under GH stimulation (1,3). Homozygous or compound heterozygous mutations in IGFALS lead to ALS deficiency. IGFALS consists of 2 exons. Until now, at least 22 different inactivating mutations of IGFALS have been reported (4-15). Similar to patients with homozygous or compound heterozygous mutations, heterozygous carriers were reported to be shorter than wild-type carriers (5,9,16,17).

We report the genotype and phenotype of a patient with ALS deficiency with a novel homozygous frameshift mutation in IGFALS presenting with short stature and reaching an adult height (AH) similar to that of heterozygous carriers of this mutation.

Methods
Molecular Studies
Informed consent was obtained from the patient and his sister and mother. The genetic analyses were performed at the Cincinnati Center for Growth Disorders, Cincinnati Children’s Hospital Medical Center. Genomic DNA was extracted from peripheral blood leukocytes.

Auxology
Height and weight were measured using a Harpenden stadiometer and electronic scale respectively and head circumference (HC) with a tape measure. Small for gestational age (SGA) was defined as birth weight and/or length SD (standard deviation) score (SDS) <−2.0. SDS for height, weight, sitting height, height SDS and HC calculated according to Turkish standards (18,19). Target height (TH) was calculated using the following equation TH = (father’s height (cm) + mother’s height (cm))/2 −6.5 cm (girls) or +6.5 cm (boys) (20). The onset of puberty was defined according to Tanner standards as attainment of testicular volume ≥4 mL in boys (21). Bone age was estimated by the Greulich and Pyle method and height prediction was calculated by Bayley-Pinneau method (22).

Serum Hormone Assays
Serum concentrations of IGF-1 and IGFBP-3 were measured by an automated immunochemiluminescence assay (Immulate 2000 XPI; Siemens Medical Solutions Diagnostics).

Case Report
A 15 3/12 year old boy was referred to the pediatric endocrinology clinic for evaluation of short stature. His height was 149.9 cm (-3.04 SDS) and his weight 52.3 kg (-2.3 SDS) with a HC 53.8 cm (-2.0 SDS) at presentation. He was 2.05 SD shorter than his TH. Clinical and laboratory characteristics of the patient at presentation and during follow-up are given in Table 1. At presentation, there were no dysmorphic features noted and no body disproportion. He was prepubertal; his testicular volumes were 3 ml bilaterally. He was born SGA at 40 weeks of gestation, with a weight of 2400 g (-2.7 SDS). Neuromotor development was normal. His medical history was otherwise unremarkable. His parents were unrelated but originated from the same village. Table 2 shows clinical and laboratory characteristics of the patient and his mother and sister. Mother’s height was 155.6 cm (-1.28 SDS). She reported achieving menarche at 13 years. Father passed away due to chronic renal failure. His reported height was approximately 170 cm (-0.2 SDS). There was no information of the father’s pubertal timing. TH of the patient was 169.3 cm (-0.99 SDS). His elder sister’s birth weight was 3000 g at 40 weeks of gestation (-0.9 SDS) and height was 157.1 cm (-1.02 SDS) at 21 years of age. Her age at menarche was 12 years. Serum IGF-1 concentration of patient [68.6 ng/mL; (normal:193-731 ng/mL)] was markedly reduced. IGFBP-3 concentration was extremely low [<0.5 ng/mL; normal: 3.2–8.7 ng/mL]. Thyroid function was normal. The peak GH response to GH stimulation test was high (31.6 ng/mL). Bone age was 13.5 years at presentation. IGFALS generation test showed a response of serum IGF-1 (from 58.4 ng/mL to 100 ng/mL), but no response of serum GH response to GH stimulation test was high (31.6 ng/mL). Bone age was calculated according to Turkish standards (18,19). Target height (TH) was calculated using the following equation TH = (father’s height (cm) + mother’s height (cm))/2 −6.5 cm (girls) or +6.5 cm (boys) (20). The onset of puberty was defined according to Tanner standards as attainment of testicular volume ≥4 mL in boys (21).

Bone age was estimated by the Greulich and Pyle method and height prediction was calculated by Bayley-Pinneau method (22).

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During follow-up fasting glucose levels of the patient were within normal range. When he was 20.1 years old, oral glucose tolerance test showed insulin resistance (Table 2). He did not have bone pain and any fracture, his spine (L1–L4) BMD determined by DXA at the age of 20.1 years was -3.5 SDS, showing osteoporosis. Serum calcium, phosphate, alkaline phosphatase, parathyroid hormone and 25-OH vitamin D levels were normal. Puberty of the patient started at 15.9 years, testes volumes were 6 mL/6 mL (Tanner stage G2). His height was -3.0 SDS at onset of puberty. His bone age was retarded by approximately 2 years when compared to his chronological age. His peak height velocity was 7 cm/year during progression of puberty (Tanner stage G3; Figure 1) and total height gain during puberty was 19.6 cm. At his last follow up visit at 20.1 years, his height SDS, weight SDS and HC SDS values were -1.08, -0.31 and -1.0, respectively. He reached his TH (Table 1). Puberty was completed and testicular volumes were 20/20 ml at age 20.1 years. His bone age was 18 years. Serum LH and FSH concentrations were 2.77 mIU/mL (normal: 1.7–
Our patient was born SGA and his birth size was significantly smaller than his heterozygous carrier sister. A deficiency may have low BMD but it was found to be normal in others (5-7,11,16,17). Reported that some cases with ALS deficiency are born SGA (7,11,12,15,16). Effect of being SGA on AH in insulin levels. Insulin resistance was reported previously in some patients with ALS deficiency (4,6,11,16). It was suggested that this may be related to the increased GH levels and the low IGF-1 levels (25).

Although our patient had pubertal delay, he demonstrated an obvious pubertal growth acceleration and good pubertal height gain and reached an AH comparable to his TH. The effect of ALS deficiency on pubertal growth pattern remains controversial. Age of pubertal onset and growth pattern are still unclear in these patients. Delayed puberty was reported in 50% of males with ALS deficiency, however, normal pubertal growth has been reported in some patients with ALS deficiency (4,6,8,10,11). An adolescent female was reported with a novel homozygous mutation of the IGFALS gene with absent pubertal growth spurt and a slow pubertal progression despite a normal onset of puberty (7). Our patient and other family members who are heterozygous carriers do not have microcephaly and their HC SDSs were similar. Microcephaly was previously reported in some patients but not present in other reported cases (5-7,9,12). It was reported that 3 siblings with an IGFALS mutation had HCs that were lower than those of heterozygous and wild-type carriers and mean HC SDS of heterozygous carriers was 0.7 SD lower than those of non-carriers. It is speculated that microcephaly may be related to the low IGF-1 levels related to the ALS deficiency during fetal life (5).

In our patient, low IGF-1, an extremely low IGFBP-3 level and moderate short stature at presentation pointed to the possibility of ALS deficiency. Molecular genetics analysis for IGFALS revealed a homozygous mutation in exon 2 (c.1663-1664delTC and p.Ser555Thrfs.19). This frameshift point mutation resulted in a substitution of a serine for a threonine at position 555 of the protein leading to an early stop codon 19 codons later (Figure 2). His mother and elder sister were heterozygous mutation carriers.

Discussion

In our patient, low IGF-1, an extremely low IGFBP-3 level and moderate short stature at presentation pointed to the possibility of ALS deficiency. Molecular genetics analysis for IGFALS revealed a homozygous mutation in exon 2 (c.1663-1664delTC and p.Ser555Thrfs.19). This frameshift point mutation caused a substitution of a serine for a threonine at position 555 of the protein leading to an early stop codon 19 codons later (Figure 2). This frameshift mutation reported in our patient could cause early protein termination and likely desaturize ALS thus leading to nonsense-mediated decay of the truncated mRNA, resulting in ALS deficiency in our patient. Serum IGF-1 and IGFBP-3 levels were not so profoundly low in our patient’s family members who were heterozygous carriers, similar to other heterozygous carriers reported previously (9,16,17). Similar to most reported cases, the main clinical feature of our patient is moderate short stature before puberty. His height SDS at presentation (-3.04 SDS) is consistent with previous reports in ALS deficient individuals (16). During follow-up, our patient showed a normal growth pattern in puberty and reached his TH in contrast with most of the cases reported in the literature in which AH was approximately 1.3 to 1.5 SD below their TH SDS (4-6,10). Phenotypic variations between patients who are homozygous for IGFALS mutation have been reported. Even a degree of phenotypic variation between two siblings was demonstrated (12). Schreiner et al (8) reported an ALS deficient patient with normal height (-0.19 SDS) and growth pattern with a difference of approximately 0.5 SDS between AH SDS and TH SDS. Although van Duyvenvoorde et al (5) reported that the sitting height/height ratio was in the upper normal range in most cases, sitting height/height ratio of our patient was normal.

AH SDS of our patient is similar to heights of his heterozygote mother and sister. The height SDSs of heterozygous carriers in our family are approximately 1 SD lower than population mean. This is consistent with previous reports indicating that heterozygous carriers are 1.0 SDS shorter than wild type subjects (5,9,17). The heterozygous carriers have a milder phenotype compared to cases with homozygous mutations (9,12,17). It is proposed that there could be a possible gene dosage effect (9,16).

Our patient and other family members who are heterozygous carriers do not have microcephaly and their HC SDSs were similar. Microcephaly was previously reported in some patients but not present in other reported cases (5-7,9,12). It was reported that 3 siblings with an IGFALS mutation had HCs that were lower than those of heterozygous and wild-type carriers and mean HC SDS of heterozygous carriers was 0.7 SD lower than those of non-carriers. It is speculated that microcephaly may be related to the low IGF-1 levels related to the ALS deficiency during fetal life (5).

Our patient showed decreased BMD Z score but BMD Z scores of heterozygous family members were within the normal range. There was no history of bone pain or fracture. Several reports suggested that patients with ALS deficiency may have low BMD but it was found to be normal in others (5-7,11,16,17). Our patient was born SGA and his birth size was significantly smaller than his heterozygous carrier sister. Although he was SGA, he reached an AH comparable to that of his sister with normal birth weight. It has been reported that some cases with ALS deficiency are born SGA (7,11,12,15,16). Effect of being SGA on AH in these patients needs to be investigated.

Although our patient had pubertal delay, he demonstrated an obvious pubertal growth acceleration and good pubertal height gain and reached an AH comparable to his TH. The effect of ALS deficiency on pubertal development remains controversial. Age of pubertal onset and growth pattern are still unclear in these patients. Delayed puberty was reported in 50% of males with ALS deficiency, however, normal pubertal growth has been reported in some patients with ALS deficiency (4,6,8,10,11). An adolescent female was reported with a novel homozygous mutation of the IGFALS gene with absent pubertal growth spurt and a slow pubertal progression despite a normal onset of puberty (7).

Our patient had insulin resistance but heterozygous carriers in this family have normal fasting glucose and insulin levels. Insulin resistance was reported previously in some patients with ALS deficiency (4,6,11,16). It was suggested that this may be related to the increased GH levels and the low IGF-1 levels (25).
In conclusion, we report a novel IGFALS mutation identified in a patient with biochemical signs of ALS deficiency and short stature, born SGA, delayed puberty but normal growth and AH comparable to TH. It is important to know all of the phenotypic features of ALS deficiency in order to ensure proper follow up of these patients. Although heterozygosity for ALS affects height, it seems to show no effect on prenatal growth, puberty, insulin metabolism and BMD.

Authorship contribution:
Concept: Sukran Poyrazoglu, Feyza Darendeliler
Design: Sukran Poyrazoglu, Feyza Darendeliler
Data Collection and Processing: All authors
Analysis and interpretations: All authors
Literature search: Sukran Poyrazoglu, Feyza Darendeliler
Writing: Sukran Poyrazoglu, Feyza Darendeliler

Financial Disclosure: The authors declare that this study received no financial support

Conflict of interest: The authors have nothing to disclose

References


**Table 1: Clinical characteristics of patient at presentation and during follow-up**

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<th>At presentation</th>
<th>At onset of puberty</th>
<th>At adult height</th>
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<tr>
<td>Age (year)</td>
<td>15.25</td>
<td>15.7</td>
<td>20.1</td>
</tr>
<tr>
<td>Height (cm) (SDS)</td>
<td>149.9 (-3.04)</td>
<td>153.2 (-2.96)</td>
<td>169.5 (-1.08)</td>
</tr>
<tr>
<td>Weight (kg) (SDS)</td>
<td>43.7 (-2.3)</td>
<td>48.2 (-2.06)</td>
<td>68.8 (-0.31)</td>
</tr>
<tr>
<td>BMI (kg/m²) (SDS)</td>
<td>19.5 (-0.72)</td>
<td>20.45 (-0.5)</td>
<td>23.9 (0.28)</td>
</tr>
<tr>
<td>Head circumference (cm)</td>
<td>54.1 (-1.8)</td>
<td>54.4 (-1.83)</td>
<td>56.2 (-1.0)</td>
</tr>
<tr>
<td>Sitting height/height</td>
<td>0.53 (0.0)</td>
<td>0.53 (-0.84)</td>
<td>0.52 (-0.6)</td>
</tr>
<tr>
<td>Testis volume (ml)</td>
<td>3/3</td>
<td>6/6</td>
<td>20/20</td>
</tr>
<tr>
<td>Bone age (years)</td>
<td>13.5</td>
<td>13.5</td>
<td>18</td>
</tr>
<tr>
<td>Target height (cm) (SDS)</td>
<td>169.3 (-1.12)</td>
<td>169.3 (-1.12)</td>
<td>169.3 (-1.12)</td>
</tr>
<tr>
<td>Predicted adult height (cm) (SDS)</td>
<td>166.2 (-1.62)</td>
<td>169.8 (-1.04)</td>
<td>170.2 (-0.97)</td>
</tr>
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</table>

**Table 2: Clinical and laboratory characteristics of the patient and heterozygous carriers at last visit**

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<tr>
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<th>Patient</th>
<th>Sister</th>
<th>Mother</th>
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<tr>
<td>Age (year)</td>
<td>20.1</td>
<td>21</td>
<td>51.3</td>
</tr>
<tr>
<td>Height (cm) (SDS)</td>
<td>169.5 (-1.08)</td>
<td>157.1 (-1.02)</td>
<td>155.6 (-1.28)</td>
</tr>
<tr>
<td>Weight (kg)</td>
<td>68.6 (-0.31)</td>
<td>49.5 (-1.48)</td>
<td>64.1 (0.88)</td>
</tr>
<tr>
<td>BMI (kg/m²)</td>
<td>24.2 (0.36)</td>
<td>20.06 (-0.94)</td>
<td>26.48 (1.75)</td>
</tr>
<tr>
<td>Head circumference (cm) (SDS)</td>
<td>56.2 (-1.0)</td>
<td>54.9 (-0.97)</td>
<td>55.1 (-0.81)</td>
</tr>
<tr>
<td>Sitting height/height (SDS)</td>
<td>0.52 (-0.6)</td>
<td>0.53 (-1.0)</td>
<td>0.54 (0.1)</td>
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5
<table>
<thead>
<tr>
<th>Parameter</th>
<th>Value 1</th>
<th>Value 2</th>
<th>Value 3</th>
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<tbody>
<tr>
<td>BMD L1–L4 Z-score</td>
<td>-3.6</td>
<td>-0.2</td>
<td>0.2</td>
</tr>
<tr>
<td>IGF-1 (ng/mL) (SDS)</td>
<td>68.6 (-2.9) (N:193-731)</td>
<td>64.7 (-3.0) (N:117-323)</td>
<td>76.8 (-1.3) (N:48.1-209)</td>
</tr>
<tr>
<td>IGFBP-3 (ng/mL) (SDS)</td>
<td>&lt;0.5 (&lt;-4.0) (N:3.2–8.7)</td>
<td>1.92 (-2.8) (N:2.9-7.3)</td>
<td>1.88 (-3.7) (N:3.4-6.8)</td>
</tr>
<tr>
<td>Fasting glucose (mg/dL)</td>
<td>77</td>
<td>77</td>
<td>98</td>
</tr>
<tr>
<td>Insulin (µU/mL)</td>
<td>27.49</td>
<td>6.4</td>
<td>9.85</td>
</tr>
<tr>
<td>Oral glucose tolerance test</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Glucose (mg/dL)</td>
<td>0' 30' 60' 90' 120'</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Insulin (µU/mL)</td>
<td>77 144 110 99 62</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>27.5 487.6 462.5 292.2 113.2</td>
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Figure 1: Growth chart of patient for height (at the upper panel) and weight (at the lower panel) plotted on Growth Chart for Turkish children (18)
Figure 2: Sequence chromatograms for patient(P), his sister(S) and mother(M). The two nucleotide deletion located immediately downstream of boxed triplex. Patient is homozygous. Sister and mother are heterozygous.

CC0251 – IGFALS c.1549_1550delTC (p.Ser517Thrfs*19)