Proniosomes: A promising approach for vesicular drug delivery

Different types of drug delivery systems are intended to deliver therapeutic agents to the appropriate site of interest to get desired pharmacological effect. In the field of drug delivery, the advancement of nanotechnology helps to develop novel dosage forms like liposome, niosome, proniosome. Proniosomes are promising drug carriers which is a dry formulation and after hydration, it converted to niosome dispersion. Dry proniosomal powder can deliver a unit dose of the drug with improved drug stability as well as solubility. By using this formulation, both the hydrophilic and lipophilic drugs can be delivered through different routes like oral, topical, transdermal, vaginal, etc. This review revolves around different features of proniosomes such as structure, formulation materials of proniosomes, preparation methods, evaluation, and application.

Keywords: Proniosomes, Vesicular drug delivery, Evaluation of proniosome, Application of proniosome

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Introduction
A novel drug delivery system is that delivers drugs at a preset rate set as per the need, pharmacologic aspects, drug profile, physiological conditions of the body, etc. In current times, there's no single drug delivery system that meets all the standards; however, efforts are created through novel approaches. One of the aims of novel drug delivery systems is targeted and controlled drug delivery. Colloidal delivery and nanotechnology have attracted the most interest as a result of promising systems having a localized result. Delivery of drugs using colloidal particulate carriers such as Proniosomes are dry, free-flowing preparation coated with a surfactant. To form a multi-lamellar niosome, Proniosomes are rehydrated directly within minutes by transient agitation. Niosome suspension is appropriate for giving medication by different routes. They are promising candidates for industrial application as they can transport, distribute, store and process easily. So, Proniosomes can be another option to
liposomal and other vesicular drug delivery systems for the entrapment of each polar and non-polar medication.² Proniosomes improve effectivity, scale back or eliminate adverse effects and enhance therapeutic actions of medicine. They are accustomed to avoid gastrointestinal tract (GIT) incompatibility, pre-systemic metabolism, and unwanted adverse effects related to oral delivery. Additionally, they maintain therapeutic levels of drug for an extended time, decrease the frequency of administration and improve patient compliance.³ ⁵

This article in brief reviews the types, fabrication, characterization, pharmaceutical, applications of Proniosomes.

**Structures of Proniosomes**

Proniosomes are microscopic lamellar structures, hexagonal structures, and blockish structures and their location is clear, semitransparent, and semi-solid gel-like structures (Figure 1). Consistent with their methodology of preparation, Proniosomes are unilamellar or multi-lamellar. They even have bilayer in their structure having hydrophilic ends that are exposed on the surface and hydrophobic chains face one another within the bilayer inside the vesicles. Bilayer consists of non-ionic surface-active agents. To create a bilayer surfactant molecule, offer direction in such a method that hydrophilic ends of the non-ionic surfactant are arranged toward the outside, whereas the hydrophobic ends exist within the opposite direction. Hydrophilic drugs are placed at intervals in the area encircled within the vesicle and also the hydrophobic medication is implanted within the bilayer. For association, in liquid media Proniosomes attach to cholesterol with different categories of non-ionic surfactant like alkyl radical or dialkyl polyglycerol ether.⁶

![Figure 01: Structure of Proniosome](image)

**Materials used for the preparation of Proniosomes**

**Surfactant**: Surfactant especially nonionic surfactant is the key structural component in the preparation of Proniosomes. These surfactants do not have any charge as they possess a polar head and non-polar tail. So, their stability, toxicity as well as compatibility is higher than other surfactants. The nonionic surfactants have wetting and emulsifying effects by which they improve the solubility and permeability of drugs. HLB value is very important for the selection of surfactants and HLB Value between 4 and 8 is compatible with vesicle formation by Proniosomes. It is difficult for hydrophilic surfactants to achieve a high concentration because of
the high liquid solubility of hydrophilic surfactants. Therefore, aggregation and conglutination to form proniosomal lamellar structure would be absent.\textsuperscript{7}

Table 1. Commonly used surfactants with their hydrophilic-lipophilic balance (HLB) values

<table>
<thead>
<tr>
<th>Name of Surfactant</th>
<th>M.W. (g/mol)</th>
<th>HLB value</th>
<th>Transition temperature</th>
</tr>
</thead>
<tbody>
<tr>
<td>Span 20 (Sorbitan monolaurate)</td>
<td>346.46</td>
<td>8.6</td>
<td>16°C</td>
</tr>
<tr>
<td>Span 40 (Sorbitan monopalmitate)</td>
<td>402.57</td>
<td>6.7</td>
<td>42°C</td>
</tr>
<tr>
<td>Span 60 (Sorbitan monostearate)</td>
<td>430.63</td>
<td>4.7</td>
<td>53°C</td>
</tr>
<tr>
<td>Tween 20 (Polyoxyethylene sorbitan monolaurate)</td>
<td>522.68</td>
<td>16.7</td>
<td>-</td>
</tr>
<tr>
<td>Tween 60 (Polyoxyethylene sorbitan monostearate)</td>
<td>648.92</td>
<td>14.9</td>
<td>-</td>
</tr>
<tr>
<td>Tween 80 (Polyoxyethylene sorbitan monooleate)</td>
<td>604.82</td>
<td>15.0</td>
<td>-</td>
</tr>
<tr>
<td>Span 85 (Sorbitan trioleate)</td>
<td>957.49</td>
<td>14.8</td>
<td>-</td>
</tr>
</tbody>
</table>

**Cholesterol:** Cholesterol can interact with nonionic surfactants and regulates the physical and structural properties of Proniosomes.\textsuperscript{8} It improves stability and rigidity of the Proniosomal membrane and controls drug permeation through the membrane. Depending on the HLB value of the surfactants, the amount of cholesterol required for the preparation of Proniosomes is determined. When the HLB value is above 10, the amount of cholesterol to be increased to cover the larger head groups.\textsuperscript{9} But, entrapment efficiency of the prepared formulation is decreased\textsuperscript{10} above a certain level of cholesterol, possibly due to a decrease in volume diameter.\textsuperscript{11}

**Lecithin:** Lecithin is a phospholipid and acts as a membrane stabilizer in the formulation of Proniosomes. The most common lecithin that is utilized in the formulation are soya and egg lecithin and it is reported that hydrogenated-type lecithins have advantages over not hydrogenated lecithins, give increased rigidity of the cholesterol and help in the formation of tight vesicles.\textsuperscript{12} Double bonds in non-hydrogenated lecithin allow the molecular chains to bend (conformational rotation), which prevents tight contact with the adjacent molecules on forming the niosomal membrane. This results in low rigidity and high permeability of the membrane.

**Hydration medium:** Generally, the hydration medium used in Proniosomes is phosphate buffer. Depending on the solubility of the encapsulated drug, the pH of the buffer is selected.\textsuperscript{13} Rukmani and Sankar\textsuperscript{14} ascertained that drug leakage was increased with the increment of the volume of hydration medium but at the same time, entrapment efficiency increases when hydration time was increased from 20 to 45 min.

**Organic Solvent:** The solvent can act as a penetration enhancer. It also greatly affects the size of vesicles formed. The size of the vesicle and permeation rate of the drug in a proniosomal formulation is influenced by the type of alcohol. Different sized vesicles are formed using different alcohols as they have the order: \( \text{isopropanol} \text{< butanol < propanol < ethanol}.\textsuperscript{15}

**Carrier material:** Carrier materials accommodate the drug in the proniosomal formulations. Carriers should have safe, non-toxicity, free-flowing properties. They should possess low solubility in the solution of loaded, but good solubility in water for ease of hydration. They increase the surface area and impart flexibility to the Proniosomes. The frequently used carrier materials are sorbitol, mannitol, maltodextrin, glucose monohydrate, spray-dried lactose, sucrose stearate, Lactose monohydrate.\textsuperscript{16

**Preparation method of Proniosomes**
A drug that has poor aqueous solubility, low bioavailability and dissolution, poor membrane permeability, low absorption profile, excessive metabolism, and variable plasma concentration, poor patient efficiency is suitable to encapsulate into Proniosomes. Three methods are available for proniosomal drug formulation. (Figure 2)

**Slurry method**

In this method, a single or mixture of organic solvent is used in the preparation of a stock solution of surfactant and membrane stabilizer. Drug and carrier are dissolved in membrane stabilizer solution and all the components are mixed until the slurry is formed. With the help of a rotary evaporator at specified conditions (e.g., 50-60 rpm, 45±2°C temperature, and 600 mm of Hg pressure), the slurry is dried, and the free-flowing product is obtained. The obtained free-flowing dried material is further dried with the help of a desiccator at room temperature under vacuum to get Proniosomes.

**Slow Spray Coating method**

The slow spray-coating method is carried out by spraying organic solution, surfactant, cholesterol, and drug onto the carrier and then removing the solvent using a rotary evaporator under controlled conditions at 65-70°C for 15-20 min. Until the desired surfactant loading has been achieved, the process is continued and repeated. The vaporization should be carried on until the powder becomes completely dry.

**Coacervation phase separation method**

Most of the proniosomal gel is prepared by this method. In this method, exactly measured amounts of drugs, surfactants, and cholesterol are placed in a clean and dry glass vial having a wide opening. Then the solvent is added and warmed in a water bath at 60-70°C until the surfactant and cholesterol are fully dissolved. To prevent the evaporation of the solvent, the open end of the vial should be covered with a lid. Followed by the addition of an aqueous phase in the vial, the mixture is warmed in the water bath to get a clear solution. Then it is cooled at room temperature, between this time proniosomal gel is produced from the dispersion.
Figure 2: Schematic representation of materials and formulation methods of Proniosomes

Evaluation of Proniosomes
A group of properties of Proniosomes can be evaluated by different methods. They are-

Vesicle size and shape
On hydration, Proniosomes are converted to globular-shaped niosomes. The size and morphology of the niosomes can be determined by optical microscopy, scanning electron microscopy (SEM), photon correlation microscopy, transmission electron microscopy (TEM), and freeze-fracture electron microscopy (FFEM). Dynamic light scattering (DLS) method is applied to measure the vesicular size distribution. DLS essentially measures fluctuations in scattered light intensity due to diffusing particles, the diffusion coefficient of the particles can be determined. The vesicle size of the niosome is measured without agitation and with agitation. If hydration is done without agitation biggest size is formed.

Angle of repose measurement
The angle of repose of dried Proniosomes prepared by slurry and spray coating method is measured by funnel and cylinder technique.

Zeta potential
The stability of the particle can be ensured with the value of zeta potential. This is ascribed to the electrostatic repulsion between particles with the same electric charge that causes segregation of the particles. High zeta potential value leads to increase repulsive interactions in charged particles and prevents the agglomerate formation between the particles. This ensures uniform size distribution in Proniosomes. Proniosomal formulation having a zeta potential value minimum ±30 mV is considered a physically stable formulation. So, aggregation of particles can be avoided.

Osmotic shock
Osmotic shock study helps in the determination of vesicle size changes. For this, the proniosomal formulations are incubated in different types of solution like hypertonic, isotonic, hypotonic solutions for 3 h. Changes in vesicle size are detected by optical microscope.\textsuperscript{23}

**Entrapment efficiency**

To study the entrapment efficiency, we need to separate the free drug by several techniques like dialysis, gel filtration, ultracentrifugation, column chromatography, freeze-thawing. Two techniques can be applied to measure entrapment efficiency. One is the destruction of the proniosomal vesicle with propane (50%) or triton (0.1%) and the entrapped drug is determined. Another method is after the destruction of the vesicle, the un-entrapped drug is measured.\textsuperscript{26}

The percentage of entrapment is calculated using the following formula:

\[
\% \text{ Entrapment} = \frac{ED}{TD} \times 100
\]

Where ED is the amount of entrapped drug and TD is the initial amount of drug.

**Drug content**

The calibration curve is used to calculate drug content. For this, Proniosomes are lysed with methanol in a volumetric flask by shaking for 15 min. Then the stock solution is prepared with methanol. With the help of phosphate buffer, 10% solution is prepared from the stock solution. Aliquots are withdrawn and absorbance is measured followed by a drawing of calibration curve.\textsuperscript{24}

**Particle size measurement**

One of the important criteria of prepared Proniosomes is particle size. With the help of scanning electron microscopy (SEM), uniformity of particle size and surface characteristics is measured. Optical microscopy is utilized to confirm the formed vesicle after hydration of Proniosome.\textsuperscript{26}

**Rate of spontaneity**

The rate of spontaneity is the measure of the number of niosomes formed following hydration of Proniosomes. To determine the rate of spontaneity, proniosomal gel is transferred and spread uniformly along the walls of the small stoppered glass tube container. Then NaCl (0.154 M) is added with caution and placed to one side without any turbulence. With the help of Neubauer’s chamber, the number of niosome eluted from Proniosomes is calculated.\textsuperscript{27}

**In-vitro dynamics study**

The in-vitro release study is a critical test to assess the safety, efficacy, and quality of nanoparticle-based drug delivery systems. In-vitro, the drug release kinetics of the prepared Proniosomes can be determined by using Franz diffusion cell, Keshary-Chien diffusion cell, dialysis membrane, reverse dialysis, United states pharmacopeia (USP) dissolution apparatus Type-1.\textsuperscript{28}

**Stability study**

To ensure the stability of the prepared proniosomes, they are placed at a various temperature like freezing temperature (2°-8°C), normal temperature (25°± 0.5°C) and elevated temperature (45°C ± 0.5°C) for 1-3 months and the change in drug content & mean vesicle diameter are observed at a different time interval. The International Conference on harmonization (ICH) guidelines
propose dry Proniosomes powder should be studied for the accelerated stability at 75% relative humidity and 40°C as per international geographical zone & geographical conditions.  

**Proniosomal drug delivery through different routes**

**Oral routes**

The oral route of drug administration is the most preferred route for drug delivery. But bioavailability of the orally administered drug is sometimes affected by first-pass metabolism, instability in the gastric environment, low permeability through the intestinal epithelium. In some cases, absorption of the drug may alter due to the presence of food. So, to improve the bioavailability of the oral drug, different nanocarriers are engaged. Oral Proniosomes are one of them that can solve the limitations of the conventional oral dosage form. In-vitro release kinetics of Oral Vinpocetine (VP) prepared by the slurry method indicated a faster release rate of reconstituted niosomes in contrast to VP suspension at pH 6.8 or 7.2 phosphate-buffered saline (PBS). In-vivo pharmacokinetic study data also showed a better correlation with the in-vitro data. Oral Acemetacin (AC) also prepared by the slurry method in proniosomal powder and tablet formulations showed better pharmacokinetic properties. Lornoxicam is a widely used analgesic drug that belongs to the Nonsteroidal anti-inflammatory group. Proniosomal form of Lornoxicam showed significantly higher (p < 0.05) transmucosal flux across the oral mucosa than Lornoxicam containing carbopol gel and the diffusion of lornoxicam was higher (more than two folds) in proniosomal formulation. Proniosomal Telmisartan tablets prepared with surfactants having different HLB values (span 40 and Brij 35), cholesterol (20-50%), and phospholipids (egg yolk and soybean). In-vitro as well as in-Vivo comparative study showed extended drug release with a higher C_{max}. The C_{max} was increased by 1.5-fold while AUC_{0-∞} also increased significantly by 3-fold when compared with the commercial tablet. The sustained release pattern of Telmisartan was indicated by t_{max} which was increased by 3-fold in contrast to conventional tablets. The relative bioavailability was also increased by 3.2-fold.

**Table 1: Proniosomal formulation delivered through different routes**

<table>
<thead>
<tr>
<th>Route</th>
<th>Preparation method</th>
<th>In-vitro/in vivo effects</th>
<th>Ref</th>
</tr>
</thead>
<tbody>
<tr>
<td>Oral Vinpocetine (VP)</td>
<td>slurry method</td>
<td>VP-loaded proniosomes produced a significant improvement in the bioavailability</td>
<td>31</td>
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<tr>
<td>Oral Acemetacin (AC)</td>
<td>slurry method</td>
<td>Both proniosomal powder and tablet formulations showed better pharmacokinetic properties.</td>
<td>32</td>
</tr>
<tr>
<td>Oral Lornoxicam</td>
<td>Proliposome-liposome method</td>
<td>The transmucosal flux of proniosomal formulations Lornoxicam across the oral mucosa was significantly higher (p &lt; 0.05) than lornoxicam containing carbopol gel and the percent drug diffused increased more than two folds.</td>
<td>33</td>
</tr>
<tr>
<td>Oral Telmisartan Tablets</td>
<td>Slurry method</td>
<td>In-vitro as well in-Vivo comparative study showed extended drug release</td>
<td>34</td>
</tr>
<tr>
<td>Ocular Lomefloxacin HCl (LXN)</td>
<td>Coacervation phase separation method</td>
<td>The result showed the area under the curve (AUC) of proniosomal formulation is higher than the marketed product</td>
<td>36</td>
</tr>
<tr>
<td>Ocular Tacrolimus</td>
<td>The proliposome-liposome method</td>
<td>Showed enhanced precorneal permeation and retention of tacrolimus</td>
<td>37</td>
</tr>
<tr>
<td>Drug/Method</td>
<td>Type/Method</td>
<td>Description</td>
<td>Reference</td>
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<tr>
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<tr>
<td>Curcumin-loaded ocular proniosomal gel</td>
<td>Coacervation-phase method</td>
<td>Proniosomal gel showed enhanced permeability</td>
<td>38</td>
</tr>
<tr>
<td>Ocular Brimonidine tartrate (BRT)</td>
<td>Coacervation phase separation method</td>
<td>The in-vivo pharmacodynamic study confirmed the improved ocular bioavailability of BRT selected formula when compared with marketed product</td>
<td>39</td>
</tr>
<tr>
<td>Ocular Dorzolamide-HCl</td>
<td>Coacervation phase separation method</td>
<td>In-vivo performance of optimized Dorzolamide-HCl showed sustained effect and a significant reduction in intraocular pressure (IOP)</td>
<td>40</td>
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<tr>
<td>Cromolyn sodium nebulizer</td>
<td>Proniosomes were prepared according to the method developed by Hu and Rhodes</td>
<td>High nebulization efficiency percentage and good physical stability</td>
<td>26</td>
</tr>
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<td>Aerosol Beclometasone dipropionate (BDP) niosomes</td>
<td>Thin-film method</td>
<td>Entrapment of BDP in proniosome-derived niosomes was higher than that in conventional thin film-made niosomes.</td>
<td>42</td>
</tr>
<tr>
<td>Vaginal Tenofovir Disoproxil Fumarate</td>
<td>Coacervation phase separation</td>
<td>The result showed a controlled and sustained release rate</td>
<td>44</td>
</tr>
<tr>
<td>Vaginal Terconazole</td>
<td></td>
<td>Results indicated that the selected formula, showed good stability and provided higher mucoadhesion and retention time then the commercial product which resulted in more efficient in-vitro inhibition of candida albicans</td>
<td>45</td>
</tr>
<tr>
<td>Parenteral Flurbiprofen</td>
<td>Slurry method</td>
<td>Sustained anti-inflammatory activity and reduce dosing frequency.</td>
<td>46</td>
</tr>
<tr>
<td>Parenteral Letrozole (LTZ)</td>
<td>Slurry method</td>
<td>Drug release exhibited a biphasic pattern, being fast at the first 24 h (up to 65% released) followed by a very slow-release phase for one month, releasing at least 95%. Overall, in this study, a facile approach to generating niosomes incorporating LTZ using a slurry-based Proniosome technology was demonstrated.</td>
<td>47</td>
</tr>
<tr>
<td>Transdermal Piroxicam</td>
<td>Coacervation phase separation method</td>
<td>Maximum flux achieved was 35.61 µg/cm²/h, an enhancement of 7.39 times was achieved for the transdermal system based on proniosomal gel as compared to control gel.</td>
<td>48</td>
</tr>
<tr>
<td>Transdermal Ketoprofen</td>
<td>Slurry method</td>
<td>Showed significantly higher cumulative amount of drug permeated and steady-state transdermal flux compared to plain gel.</td>
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<td>--------------------------------------------------------------------------------------------------</td>
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<tr>
<td>Transdermal Meloxicam</td>
<td>Coacervation phase separation method</td>
<td>The prepared proniosomes significantly improved drug permeation and reduced the lag time ($p&lt;0.05$). Proniosomes prepared with Span 60 provided a higher meloxicam flux across the rat skin than did those prepared with Tween 80. Testing of the anti-inflammatory effect of meloxicam proniosomal gel showed better pharmacological activity when compared with the standard meloxicam gel.</td>
<td></td>
</tr>
<tr>
<td>Transdermal Celecoxib</td>
<td>Coacervation phase separation method</td>
<td>The selected proniosomal gel produced 100% inhibition of paw edema in rats up to 8 h after carrageenan injection. It produced 95% and 92% inhibition after 12 h and 24 h, respectively.</td>
<td></td>
</tr>
<tr>
<td>Transdermal Tenoxicam</td>
<td>Coacervation phase separation method</td>
<td>The investigated tenoxicam loaded proniosomal formula proved to be non-irritant, with significantly higher anti-inflammatory and analgesic effects compared to that of the oral market tenoxicam tablets.</td>
<td></td>
</tr>
<tr>
<td>Transdermal Fluconazole</td>
<td>Coacervation phase separation method</td>
<td>The results showed a well-defined spherical vesicle with sharp boundaries. Moreover, this formulation showed an excellent microbiological activity represented by a greater zone of inhibition (5.3 cm).</td>
<td></td>
</tr>
<tr>
<td>Transdermal Estradiol</td>
<td>Coacervation phase separation method</td>
<td>The Pronosome gel provided a higher permeation flux of estradiol across the skin as compared to the niosome suspension.</td>
<td></td>
</tr>
<tr>
<td>Dermal Boswellic acid</td>
<td>Coacervation phase separation method</td>
<td><em>In-vitro</em>, skin permeation study showed the most sustained release in 24 h.</td>
<td></td>
</tr>
<tr>
<td>Transdermal Atorvastatin Calcium (ATC)</td>
<td>Coacervation phase separation method</td>
<td>Results suggested a promising, easy-to-manufacture, and effective ATC proniosomal gel for safe treatment of hyperlipidemia</td>
<td></td>
</tr>
<tr>
<td>Transdermal Cilostazole (CLZ)</td>
<td>Coacervation phase separation method</td>
<td>The CLZ-loaded proniosomes showed promising results with high potential to deliver it across the skin.</td>
<td></td>
</tr>
<tr>
<td>Transdermal Clozapine (CLZ)</td>
<td>Coacervation phase separation method</td>
<td>The results suggested that clozapine could be effectively loaded into proniosomal gel for administration through skin.</td>
<td></td>
</tr>
<tr>
<td>Transdermal Coacervation phase</td>
<td>The in vitro drug diffusion studies revealed</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Ocular routes

Proniosomes are one of the promising methods in ocular drug delivery. In this route, proniosomal gels in the ocular route provide several advantages like extended and sustained action, enhanced corneal residence time, prevent enzymatic degradation of drugs in tears, and ultimately improve ocular bioavailability. Lomefloxacin HCl (LXN) prepared by the coacervation phase separation method. The result showed the area under the curve (AUC) of proniosomal formulation is higher (722.45 ± 0.01) than the marketed product (126.25 ± 0.049). Li et al. developed Proniosomal formulation of Tacrolimus, an immunosuppressive agent for topical ophthalmic delivery containing lecithin and poloxamer 188 as surfactants, cholesterol. In-vitro studies in rabbit cornea of Stable tacrolimus-loaded proniosomes showed enhanced precorneal permeation and retention of tacrolimus. The in-vivo ocular irritation test of 0.1% tacrolimus-loaded proniosomes in rat eyes for 21 successive days (four times in a day) showed no irritation and good compatibility with the cornea. Proniosomes were also found to prolong the survival of corneal grafts and showed practical corneal anti-allograft rejection efficacy in the xenotransplantation model. Curcumin-loaded proniosomal gel was formulated for the treatment of ocular inflammation with the help of Cremophore RH, lecithin, and cholesterol. Selected proniosomal gel showed 3.22-fold and 1.76-fold higher permeability rates than Curcumin dispersion and its freeze-dried form respectively. This formulation could be an effective, better biofriendly alternative for the therapy of inflaming eyes.

Emad Eldeeb et al. studied on Brimonidine tartrate (BRT) Proniosomal formulation used in glaucoma developed by coacervation phase separation method. They utilized two surfactants having higher and lower HLB values namely Tween 80 (HLB 15) and Span 60 (HLB 4.7) respectively. The result of BRT Proniosomal gel showed a 7.90-fold increase in mean residence time (MRT) in comparison to the marketed product. At the same time, the relative area under the plasma concentration-time curve over the last 24-h dosing interval (AUC0-24) value was also increased to 5.024-folds. Draize test also ensured that the formulations had no ocular irritation. Dorzolamide-HCl (Dorz) is an anti-glaucoma drug soluble in water. The proniosomal gel of this drug was formulated to sustain its effect and to reduce dosing frequency. Here the gel was formulated by utilizing L-α -lecithin, Cholesterol, span 40 using the coacervation phase separation method. The in-vivo results showed a maximum reduction of intraocular pressure (IOP) was 32.6 ± 2.7 at 1.5 h. On contrary, the proniosomal gel showed a sustained decrease in IOP with a maximum value of 45.4 ± 8.2 at 6 h which was significantly higher than standard formulation. Even after completion of the experimental period (8 hours), the % dec in IOP was 19.5 ± 9.2. This implies the prolonged release of Dorzolamide-HCl proniosomal gel from the optimized formulation.

Pulmonary routes

<table>
<thead>
<tr>
<th>Galantamine hydrobromide separation method</th>
<th>that the proniosomal gel containing Tween 80 showed maximum drug diffusion (99.24%) and the gel containing Span 20 showed minimum drug diffusion (71.74%).</th>
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<tbody>
<tr>
<td>Intrasinal Duloxetine Coacervation phase separation method</td>
<td>Result showed significantly improved permeation enhancement and stability with better control over drug release for a longer period through intranasal administration</td>
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**Ocular routes**

Proniosomes are one of the promising methods in ocular drug delivery. In this route, proniosomal gels in the ocular route provide several advantages like extended and sustained action, enhanced corneal residence time, prevent enzymatic degradation of drugs in tears, and ultimately improve ocular bioavailability. Lomefloxacin HCl (LXN) prepared by the coacervation phase separation method. The result showed the area under the curve (AUC) of proniosomal formulation is higher (722.45 ± 0.01) than the marketed product (126.25 ± 0.049). Li et al. developed Proniosomal formulation of Tacrolimus, an immunosuppressive agent for topical ophthalmic delivery containing lecithin and poloxamer 188 as surfactants, cholesterol. In-vitro studies in rabbit cornea of Stable tacrolimus-loaded proniosomes showed enhanced precorneal permeation and retention of tacrolimus. The in-vivo ocular irritation test of 0.1% tacrolimus-loaded proniosomes in rat eyes for 21 successive days (four times in a day) showed no irritation and good compatibility with the cornea. Proniosomes were also found to prolong the survival of corneal grafts and showed practical corneal anti-allograft rejection efficacy in the xenotransplantation model. Curcumin-loaded proniosomal gel was formulated for the treatment of ocular inflammation with the help of Cremophore RH, lecithin, and cholesterol. Selected proniosomal gel showed 3.22-fold and 1.76-fold higher permeability rates than Curcumin dispersion and its freeze-dried form respectively. This formulation could be an effective, better biofriendly alternative for the therapy of inflaming eyes.

Emad Eldeeb et al. studied on Brimonidine tartrate (BRT) Proniosomal formulation used in glaucoma developed by coacervation phase separation method. They utilized two surfactants having higher and lower HLB values namely Tween 80 (HLB 15) and Span 60 (HLB 4.7) respectively. The result of BRT Proniosomal gel showed a 7.90-fold increase in mean residence time (MRT) in comparison to the marketed product. At the same time, the relative area under the plasma concentration-time curve over the last 24-h dosing interval (AUC0-24) value was also increased to 5.024-folds. Draize test also ensured that the formulations had no ocular irritation. Dorzolamide-HCl (Dorz) is an anti-glaucoma drug soluble in water. The proniosomal gel of this drug was formulated to sustain its effect and to reduce dosing frequency. Here the gel was formulated by utilizing L-α -lecithin, Cholesterol, span 40 using the coacervation phase separation method. The in-vivo results showed a maximum reduction of intraocular pressure (IOP) was 32.6 ± 2.7 at 1.5 h. On contrary, the proniosomal gel showed a sustained decrease in IOP with a maximum value of 45.4 ± 8.2 at 6 h which was significantly higher than standard formulation. Even after completion of the experimental period (8 hours), the % dec in IOP was 19.5 ± 9.2. This implies the prolonged release of Dorzolamide-HCl proniosomal gel from the optimized formulation.
With the aid of the pulmonary route, we can easily treat respiratory diseases than other delivery methods. Through this route, drugs can be applied directly within the lungs. Drug-loaded particles like liposomes dispensed through aerosol, can easily distribute to bronchi and lungs and prolong the release of the drug. Liposomal delivery also has minimum systemic side effects due to localized action to the lungs. But liposomes may be degraded by oxidation or hydrolysis. So, the proniosome can be an option to overcome the limitations of the liposome. For pulmonary drug delivery, the Air-jet nebulizer is known very well. proniosome-derived niosomes of cromolyn sodium were prepared by AbdElbary et al. They used sucrose stearates in the formulation. The result showed a controlled release of drugs from the proniosome-derived niosomes compared to standard drug solution. Furthermore, high nebulization efficiency and physical stability were also achieved.

Likewise, aerosol properties of beclometasone dipropionate (BDP) niosomes using Aeroneb Pro and Omron Micro Air vibrating mesh nebulizers and Pari LC Sprint air-jet nebulizer was investigated by Elhissi et al. The study showed the satisfactory entrapment efficiency of BDP in proniosome-derived niosomes and the value was higher than that in conventional thin film-made niosomes.

**Vaginal routes**

Vaginal drug delivery is one of the favorable routes to target the disease associated with women's health issues. It offers both the local and systemic delivery of drugs. Usually, different categories of drugs like antibiotics, antifungal, antiprotozoal, antiviral, labor-inducing agents, spermicidal agents, steroids, etc. are delivered through the vaginal route. Proniosomal gel has very good mucoadhesive properties and provides a constant release pattern, which is very useful for vaginal drug delivery. Tenofovir Disoproxil Fumarate (TDF) is an antiretroviral drug (nucleotide analog) that works through the inhibition of viral reverse transcriptase. Proniosomal gel of TDF was prepared with the help of Cholesterol, surfactants (Span 20, 40, 60, 80, Tween 20, Tween 80), Lecithin by coacervation phase separation method. A comparative in-vivo dissolution study had been conducted between proniosomes suppository, Drug suppository, and proniosomal gel formulations for 24h using cellophane membrane, our results showed that the proniosomal suppository. The result showed a controlled and sustained release rate when compared to the other two formulations. Terconazole, antifungal drug, proniosomal gels were developed based on span 60 and Brij 76 in different molar ratios (1:1, 1:1.5 and 1:2) relative to cholesterol. The result showed that increased concentration of cholesterol relative to surfactant has affected both Entrapment Efficiency and vesicle size. niosomes prepared by incorporating into 1% carbopol gel. Drug release profile from different prepared proniosomal gel formulations in simulated vaginal fluid (SVF) was studied in comparison to the commercial product of terconazole for 24 hours. Depending on the high EE% and in-vitro release profile, selected formulation was further evaluated for stability, mucoadhesion to the vaginal mucosa and inhibition of candidas growth. Results indicated that the selected formula, showed good stability and provided higher mucoadhesion and retention time then the commercial product which resulted in more efficient in-vitro inhibition of candida albicans.

**Parenteral routes**

In parenteral drug delivery, targeted and sustained drug release at a predetermined rate can be achieved due to remarkable advancement in pharmaceutical technology. Flurbiprofen and Letrozole (LTZ) both are prepared by the slurry method. Both drugs showed sustained activity and reduced dosing frequency.

**Dermal and transdermal routes**
The dermal route is employed for local action only to treat different types of skin disease. This route can avoid systemic effects and therefore offers fewer side effects. On the other hand, through transdermal delivery, we can deliver drugs for systemic action. But in both the dermal and transdermal drug delivery, the skin prevents the penetration of drugs. The vesicular drug delivery can be utilized to overcome the problem.

Nonsteroidal anti-inflammatory drugs (NSAIDs) like Piroxicam\textsuperscript{48} Ketoprofen\textsuperscript{49}, Meloxicam\textsuperscript{50}, Celecoxib\textsuperscript{51}, Tenoxicam\textsuperscript{30} are formulated to avoid gastrointestinal adverse effects. Here all the NSAIDs except Ketoprofen is prepared by coacervation phase separation method, whereas Ketoprofen is prepared by the slurry method.

Fluconazole-loaded proniosomal gels were prepared by the coacervation phase separation method using different nonionic surfactants (spans and tweens). The prepared fluconazole proniosomal gels were evaluated for various parameters such as particle size (PS), drug entrapment efficiency percentage (EE%), and \textit{in vitro} drug release. The experimental results showed that the EE% for the prepared formulae are acceptable (85.14\%–97.66\%) and they are nanosized (19.8–50.1 nm). The formulated gel also showed sustain drug release. Formulation, which was prepared from span 60, tween 80 (1:1), and cholesterol showed highest EE% and gave slow release (40.50\% ± 1.50\% after 6 h), was subjected to zeta potential (ZP) test, transmission electron microscopy as well as microbiological study. The results showed a well-defined spherical vesicle with sharp boundaries with good physical stability of fluconazole within the prepared gel. Moreover, this formulation showed an excellent microbiological activity represented by a greater zone of inhibition (5.3 cm) compared to control gel (fluconazole in 2\% hydroxy propyl methyl cellulose (HPMC) gel formula) (4.2 cm) and plain gel with no drug (0 cm) against Candida albicans.\textsuperscript{52} Fang et al\textsuperscript{53} studied transdermal Estradiol gel and the result provided a higher permeation flux of estradiol across the skin. \textit{In-vitro} skin permeation study of dermal Boswellic acid gel, was studied for 24 hours, and a sustained release pattern was observed (84.83 ± 0.153 mg/cm\textsuperscript{2}). Inhibition of inflammation of the proniosomal patch was also significantly ((p < 0.001) higher compared to the marketed gel at the same dose.\textsuperscript{54}

In an attempt to modify the anti-hyperlipidemic effect and to reduce statins-induced hepatotoxicity, Atorvastatin Calcium (ATC) transdermal proniosomal gel (PNG) was developed by coacervation phase separation method. Different non-ionic surfactants (Spans, Tweens, Cremophor RH 40, and Brij 52) were incorporated in the vesicle's lipid bilayer, in combination with lecithin. PNG gel was characterized for encapsulation efficiency percent (% EE), vesicle size, polydispersity index (PDI), and zeta potential (ZP). The results revealed nano-size (≤350 nm) range vesicles with relatively high ATC entrapment efficiency (70.12–88\%). \textit{Ex-vivo} results of the selected formulation demonstrated the permeation superiority of ATC proniosomes over free drugs. The selected proniosomal gels showed significantly high flux ranging from 4.23 to 8.46 µg/cm\textsuperscript{2} h\textsuperscript{−1} with permeability coefficient values (P) (0.004–0.008 cm/h) when compared to free ATC dispersion which significantly possessed lower flux and permeability coefficient results (2.92 µg/cm\textsuperscript{2} h\textsuperscript{−1} and 0.003 cm/h respectively). The pharmacodynamic study revealed that transdermal administration of ATC- PNG succeeded in retaining the antihyperlipidemic efficacy of orally administered ATC without elevating liver biomarkers. The histological examination signified the role of optimized ATC-PNG in hindering statin-induced hepatocellular damage.\textsuperscript{55} Transdermal Cilostazole (CLZ) proniosomes were prepared by a coacervation phase separation technique. The optimum formula(opt) composed of 540 mg Span60 and 59.7 mg of cholesterol, had the highest entrapment efficiency (EE%) of (75.12 ± 0.125\%), particle size (PS) of (300.3 ± 0.2 nm), zeta potential (ZP) of (−39.35 ± 0.15 mV), the percent of the drug...
released after 2h was (24.32 ± 0.13%) and after 24h was (81.175 ± 0.325%). The Safety of the Proniosomes for topical application was confirmed by the histopathological examination. The CLZ-loaded proniosomes showed promising results with high potential to deliver it across the skin. Clozapine (CLZ) loaded proniosomal gel (PN) gel was prepared by the coacervation phase separation method utilizing span-60, cholesterol and lecithin. The optimized formulation had the highest entrapment efficiency of 90% and the average particle size of approximately 325 nm. polydispersity index (PDI) reflected homogeneity in the formulation. zeta potential (ZP) was -59.76 mV, high enough to indicate a stable formulation. The in vitro release studies manifested a sustained release behavior of clozapine from the proniosomal gel. The ex vivo permeation showed noteworthy permeation of the drug through stratum corneum with a steady state flux of 18.26 ug/cm2/hr. Galantamine hydrobromide is used for the treatment of Alzheimer's disease. Galantamine hydrobromide is formulated as proniosome gel by Coacervation phase separation method to overcome the side effects of oral delivery. Microscopical observations of the gels showed vesicles of optimum size from 3.030 mm - 3.735 mm. The gel also showed optimum rate of spontaneity in the range 9.60 mm3x1000 to 11.80 mm3x1000 and entrapment efficiency of vesicles in the range 66.15% to 86.92%. The gels had pH in suitable range of skin (5.92-6.9). The in vitro drug diffusion studies revealed that the proniosomal gel containing Tween 80 showed maximum drug diffusion (99.24%) and the gel containing Span 20 showed minimum drug diffusion (71.74%).

**Intranasal routes**

The nasal drug delivery method has some limitations like mucociliary clearance, degradation of drugs by the enzyme. Vesicular drug delivery systems can circumvent these limitations. Duloxetine (DX) is a new norepinephrine (NE) reuptake inhibitor (SNRI) used in the treatment of depression. But it has high first-pass metabolism and low bioavailability (< 50%) following oral administration eventually leading to low cerebrospinal fluid (CSF) concentrations. Khatoon et al designed mucoadhesive thiolated chitosan (TCS) gel containing proniosomes of Duloxetine (DX) for the intranasal drug delivery to enhance the drug's contact time with nasal mucosa, bypass the first-pass effect and target the brain possibly using the olfactory pathway. Here soya lecithin, cholesterol, Tween 80 were used in the preparation of the gel. The pH of the Duloxetine-loaded proniosomal gel (D-MPNG) was 5.67 ± 0.145, indicating the compatibility of formulations within the nasal cavity without producing irritation. Notably, the D-MPNG showed better control, releasing only 24% DX at pH 7.4 over 24 h compared to 78% release at pH 5.5. The presence of thiol groups of TCS significantly controls the water uptake, resulting in moderate swelling and higher viscosity; thus, providing a sustained effect for a longer period.

**Cosmeceuticals Application of Proniosomes**

Cosmeceuticals is generally used to refer to skincare products that contain active ingredients that are beneficial to improving skin's appearance and promoting healthy skin. Anti-aging cosmeceuticals are most frequently recommended by physicians who use them as an integral part of a comprehensive skin rejuvenation program. Moisturizers and serums containing ingredients like vitamin C, niacinamide, retinol, peptides, growth factors, and botanicals can all be used in this regard. In addition, patients undergoing cosmetic procedures such as laser resurfacing and chemical peels may be given cosmeceuticals to "prime" the skin for procedures, encourage healing, and reduce complications after. Cosmeceuticals are also recommended for patients with acne, rosacea, eczema, and other skin conditions where they are commonly used in combination with prescription medications. For example, moisturizers containing anti-inflammatory botanical ingredients may be used in conjunction with prescription medications for treating rosacea.
Cosmeceuticals containing soy can be used to provide added skin lightening benefits when paired with hydroquinone.
Applying therapeutic and cosmetic agents onto or through skin requires a non-toxic, dermatologically acceptable carrier, which not only controls the release of the agent for prolonging action but also enhances the penetration to the skin layer. Proniosome gel meets such criteria which are useful for delivery of cosmetics and Cosmeceuticals. The therapeutic agents which can be utilized for incorporation into proniosomal carrier systems include, moisturizing, nutritional, anti-wrinkle, anti-aging, cleansing, sunscreen particles, etc. Proniosomes are a potentially scalable method for producing niosomes for the delivery of hydrophobic or amphiphilic drugs. Anti-aging cream containing the methanolic purple glutinous rice extract loaded in niosomes was formulated by Manosroi et al. Anthocyanin present in purple glutinous rice extract is responsible for the anti-aging activity. After 6 cycles of heating and cooling test, the formulation with 1% w/v of the purple glutinous rice extract contains 52.28% anthocyanin of the initial. For in vivo antiaging activities, a cream containing niosomes loaded with the extract gave significantly decreased melanin index and skin roughness reduction of 14.05 and 9.95% of the initial, respectively. The % changes of the increased skin hydration, skin elastic extension, and skin elastic recovery when applied on human volunteers' skin with this formulation were +48.73, 24.51, and +35.98%, respectively.

Tretinoin (TRT) is a widely used retinoid for the topical treatment of acne, photo-aged skin, psoriasis, and skin cancer. TRT-loaded Proniosomes were prepared by slurry method with the help of Span 60 and D-Sorbitol, Span 40, Cholesterol 95% stabilized, and Tween 20. Prepared hydrated proniosome was characterized by evaluation particle size, the effect of drug concentration, entrapment efficiency, etc. The entrapment efficiency of all hydrated proniosomal dispersions ranged from 76.6% ± 0.001 prepared using Span 40 to 94.15% ± 0.041 prepared using Span 60. In-Vitro drug release was increased till the 5th hour.

Diferuloylmethane or curcumin is obtained from turmeric, which possesses inflammatory properties blocking the formation of reactive oxygen species. Proniosomes of curcumin were prepared using nonionic surfactants (tween 80, span 60) either solely or in combinations with cholesterol. The highest encapsulation efficiency of curcumin in niosomal formulations was 99.74%. Kinetically, niosomes fitted to the Korsmeyer-Peppas model with non-Fickian transport. The anti-inflammatory activity of curcumin in various formulations was evaluated using a rat hind paw edema method and the % of swelling was 17.5% following 24h in the group treated with curcumin niosomal emulgel.

Coenzyme Q10 (CoQ10) also known as ubiquinone, an essential compound is found in every cell of the human body. Coenzyme Q10 (CoQ10), an essential compound of cellular bioenergetics also acts as a strong antioxidant and protects the body against aging. Coenzyme Q10 proniosomes were prepared by using the standard method with the help of Q10, Span 85, soya lecithin, and Cholesterol. In-vitro drug release of CoQ10 followed a special cubic model, as the statistics of the chosen model were found significant (p= 0.0006). Change in the levels of soy lecithin had a great impact and showed a synergistic effect on the release characteristics. Skin permeation study showed the cumulative amount of CoQ10 permeated in 12 h was found to be 315.85, 463.25, and 507.49 mg/cm², respectively for selected Proniosomal gel (PN) formulations. Animal skin was treated with UV radiations followed by treatment of PN gel CoQ10 and conventional CoQ10 present in a gel base. The effectiveness of the treatment was evaluated based on biochemical estimation and histopathological studies. By using CoQ10 PN gel formulation, levels of superoxide dismutase (SOD), catalase (CAT), glutathione (GSH), and...
total proteins were restored by 81.3%, 72.1%, 74.8, and 77.1% respectively to that of a control group. Histopathological studies revealed better protection of skin treated with CoQ10 PN gel compared to free CoQ10. Prepared PN gel did not interact with the normal histology hence, tolerated by animal skin compare to conventional gel. Assessments of the formulations for various Enzymatic and non-enzymatic estimation in animal skin after UV irradiations proved the efficacy of the developed formulation.

Proniosomal formulations containing the natural antioxidant Resveratrol (RSV) were prepared by M. Schlich et al. Resveratrol (trans-3,5,4'-trihydroxystilbene, RSV) is a polyphenol compound having anti-inflammatory, neuro-protective, anti-aging, and anti-cancer effects. Proniosomal powders were prepared by the slurry method and characterized. The hydration and sonication of proniosomes resulted in the formation of lipid nanoparticles with a mean diameter in the range 180-300 nm and a highly negative surface charge. The RSV release from proniosome-derived niosomes was investigated in simulated gastric and intestinal fluid. Biocompatibility assay carried out on intestinal cells (Caco2) demonstrated that proniosomes prepared with an HLB of 13.5 showed to be significantly less toxic than their HLB16.7 counterpart. All the tested formulations could be employed safely at the doses commonly administrated by oral route.

Canthaxanthin (CTX) is a xanthophyll (a subclass of carotenoids) with widespread applications in the pharmaceutical, cosmetic. It is a superior antioxidant and scavenger of free radicals as compared with carotenoids such as β-carotene. Canthaxanthin (CTX) was encapsulated in proniosome powders. Proniosomes were prepared with an equimolar ratio of span 60/cholesterol and four different carriers, namely maltodextrin, mannitol, lactose, and pullulan. The study showed that the niosomes produced by hydration and sonication of the proniosomes were small in size (≤ 200 nm) and quite homogeneously dispersed (PDI ≤ 0.3). The encapsulation efficiency of CTX in formulations varied between 55.3±1.8% and 74.1±2.7% after hydration and sonication. Although light and high temperatures affected the stability of CTX drastically, encapsulation in proniosomes retarded its degradation. This formulation can provide convenient, nontoxic, and inexpensive vehicles for dissolving and stabilizing CTX in functional food products.

O-Padimate is a UV-B Filter widely used as a sunscreen agent. This study aimed to investigate the combined influence of 3 independent variables in the preparation of O-Padimate Proniosomes were prepared by the slurry method with Span: Brij, surfactant. The formulated gels were characterized for Vesicle size, morphology & Encapsulation efficiency, Skin permeation studies, Rheological properties, and Sun Protection Factor (SPF). Results reveal that optimized O-Padimate Proniosomal formulations showing high SPF and low TWEL (Transdermal water loss). Rutin (Rut) is a natural flavonol that has a wide range of therapeutic properties including antioxidant and antitumor activities. Rutin proniosomal gel for cutaneous applications was designed to improve the poor aqueous solubility of rutin. The gel was prepared by coacervation phase-separation method and complies with the standard requirements in terms of particle size (140.5 ± 2.56 nm), zeta potential (-27.33 ± 0.09 mV), encapsulation capacity (> 50%), pH (7.002 ± 0.18), and rheological properties. The results showed high biocompatibility of the gel on the 3D reconstructed human epidermis model characterized by increased viability of the cells and a lack of irritant and phototoxic potential. The evaluations on 2D cells confirm the preferential cytotoxic effect of Rut on melanoma cells (IC\textsubscript{50} value = 8.601 µM, nuclear fragmentation).
compared to normal keratinocytes. Our data suggest that the proniosomal gel is a promising drug carrier for Rut in the management and prevention of skin disorders.\textsuperscript{76}

**Clinical trials with Proniosomes**

Selected proniosomal formulation of vinpocetine was tested \textit{in-vivo} to compare the pharmacokinetics of vinpocetine from a proniosomal patch containing vinpocetine (treatment A) to an oral commercial tablet containing the same dose of vinpocetine (treatment B) using a non-blind, two treatment, two-period, randomized, crossover design.\textsuperscript{77} Twelve healthy non-smoking male volunteers (26–37 years, 78–96 kg) participated in the study and were randomly assigned to one of the two treatment groups of equal size. The study result showed significant differences in the shape of the concentration-time courses between the two treatments. For Vinpocetine oral tablet a rapid sharp peak was found at 1.5 hours followed by a fast decline in plasma drug levels. On the other hand, for the vinpocetine proniosomal patch, the absorption was much slower and extended over a longer period. Moreover, the patch exhibited higher drug levels in the plasma from 6 to 12 h compared to the tablet. The average $C_{\text{max}}$ was significantly lower ($p < 0.05$) for the patch (12.44 ± 1.87 ng/ml) compared with the oral tablet (63.69 ± 8.32 ng/ml) while $t_{\text{max}}$ was significantly higher ($p < 0.05$) in the case of the transdermal patch (12 h) compared with the oral tablet (1.5 h). The extent of absorption of vinpocetine from the patch expressed by $\text{AUC}_{0-t}$ was determined to be about 101% larger and statistically significantly different when compared to the oral tablet. The relative bioavailability of vinpocetine proniosomal patch to the oral tablet was estimated to be on average 206%. The elimination half-lives of vinpocetine after oral and transdermal administration were 1.36 ± 0.27 h and 13.94 ± 1.2 h, respectively, and were statistically significant ($p < 0.01$).\textsuperscript{77}

Tretinoin (TRT) loaded Proniosomes was prepared by slurry method with the help of Span 60 and D-Sorbitol, Span 40, Cholesterol 95% stabilized, and Tween 20. The formulated proniosomal gel was studied clinically on 12 Egyptian patients aged >18 years (2 males and 10 females; with an average of 20 (± 4 years) with acne (papules, closed comedones, and open comedones) on their face.\textsuperscript{64} The result showed only very slight erythema (score = 0.143 ± 0.377) for TRT proniosomal gel in comparison to 0.025 % TRT gel (score = 1.70 ± 0.755). The similarly marketed product showed an erythema score of 1.50 ± 0.534 and was not able to diminish the irritation caused by topical application of TRT. Overall improvement of the individual lesions was also better than the marketed product during the 4 weeks study period.

Niosomal (Hydrated form of proniosome) Benzoyl Peroxide (BPO) and Clindamycin (CL) Lotion were prepared and compared with Niosomal Clindamycin in acne vulgaris. In both cases, the concentration of the drug was 1%. A double-blind clinical trial study on 100 patients with acne vulgaris was conducted in Afzalipour hospital in Kerman.\textsuperscript{72} The efficacy of treatment protocols was evaluated in the 2nd, 4th, 8th, and 12th weeks of treatment by counting lesions (severity and grading acne lesions) and quality of life (QoL). Furthermore, the side effect was evaluated at each treatment visits. The reduction in mean percentage of acne lesions in the case group (treated with BPO 1% and CL1%) (64.21%) was higher than the control group (treated with niosomal CL 1%) (59.04%), but the statistical difference was not significant. A sum of excellent and good results was found in 80% and 76.1% of the case and control groups, respectively ($P=0.377$). Also adding BPO to the treatment formulation in the case group did not increase adverse effects, as the statistical difference between the 2 groups was not significant.\textsuperscript{78}
## Patents on Proniosomes

### Brief description of Patents on Proniosomes

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<td>CN103340823A</td>
<td>Liu Qiang, Zhang Bin, Liu Li, Zhu Hongxia, Li Shasha, Jiang Xiao</td>
<td>Formulation of paenol proniosomes and preparing method thereof</td>
<td>Paenol proniosomes are an effective method for solving the problems of easy aggregation and easy fusion of niosomes and leakage of drugs in a solution state and a storage process of the non-ionic surfactant niosomes. The paenol proniosomes can be directly applied to skins, and also can be prepared into a gel agent, an ointment, a patch and other transdermal drug delivery formulations.</td>
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<td>1288/DEL/2012</td>
<td>Kiran Yadav, Deepak Yadav, Sanju Nanda, Kamal Saroha</td>
<td>Curcumin proniosomal/niosomal formulation, method for its preparation and use thereof</td>
<td>Niosomal/proniosomal vesicular system can improve the dispersibility of curcumin and thus providing advantage to make curcumin in administrable or applicable form for its cosmetic/therapeutic/medical purposes.</td>
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<td>3228/DEL/2012</td>
<td>Munish Garg, Monika Joon</td>
<td>Novel Proniosomal Gel of <em>Withania Somnifera</em></td>
<td>This invention relates to preparation of proniosomal gel of standardised <em>Withania somnifera</em> leaves extract, for better bioavailability, anti-inflammatory activity, to be used as transdermal application.</td>
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<td>Munish Garg, Parul Garg</td>
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<td>Alain Ribier Jean-Thierry Simonnet</td>
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<td>Rose M. Handjani Alain Ribier Guy Vanlerberghe Arlette Zabotto Jacqueline Griat</td>
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<td>CN105311638A</td>
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<td>WO2000042987</td>
<td>Amarjit Singh</td>
<td>Targeted vesicular constructs for</td>
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| KR20040058196 | CHO, Young, W. Lee, Keith, Kwang-Ho | Pro-micelle pharmaceutical compositions | The present invention provides a composition containing a pharmaceutically active drug encapsulated with an esterified C₅-C₁₈ fatty acid membrane. The result of the conversion of the micelles (pro-micelle) - are pharmaceutically that effectively deliver the active drug in systemic circulation procedure. |

EG24388A

| Hanan Hosny Ellaithy | Ahmed Abd Elbary Ebd Elrahman Mina Ibrahim Tadros | A method for the preparation of nebulizable micronized niosomes of cromolyn sodium using nonionic surfactants | This invention involves the controlling the rate of drug release after inhalation so that biological half-life could be increased, consequently number of doses (4 - 6 daily) should be reduced. |

**Conclusion**

Proniosomes can be a promising vesicular drug delivery method for the future. They are one of the drug carriers in vesicular drug delivery methods which is a better alternative to liposomal drug delivery due to its controlled and sustained action and provide better physical, chemical stability and potentially scalable for commercial viability. They offer excellent potential for improved drug delivery, through versatile routes such as oral, parenteral, dermal, transdermal, ocular, vaginal, pulmonary, and nasal by overcoming the permeation barriers faced by several drugs. Different types of unit dosages form like tablets, capsules, and beads can be prepared with the dry form of proniosomes. Due to the versatility of proniosomes, they are widely investigated as drug carriers. There are a lot of scopes to investigate new carrier material for the preparation of proniosomes and their potential remains to be investigated to the full extent.

**References**


22.


79. Qiang L, Bin Z, Li L, Hongxia Z, Shasha L, Xiao J. Formulation of paeonol proniosomes and preparing method thereof. CN103340823A


83. Ribier A., Jean-thierry S. Process for the stabilization of vesicles of amphiphilic lipid(s) and composition for topical application containing the said stabilized vesicles. U.S. Patent US 6051250A


86. Singh A, Jain R. Targeted vesicular constructs for cytoprotection and treatment of H. pylori infections. WO2000042987A8

87. Young C, Lee W, Keith, Kwang-Ho. Pro-micelle pharmaceutical compositions. KR20040058196A

88. Abd-Elbary A, El-laithy HM, Tadros MI. A method for the preparation of nebulizable micronized niosomes of cromolyn sodium using nonionic surfactants. EG24388A