

Piroksikamın fizikokimyasal özelliklerinin belirlenmesi

Mustafa Çelebier¹, Merve Nenni², OZAN KAPLAN¹, emrah akgeyik³, Mustafa Sinan Kaynak⁴, Selma Şahin⁵

¹Hacettepe Üniversitesi Eczacılık Fakültesi Analitik Kimya Ana Bilim Dalı Ankara

²Çukurova Üniversitesi Eczacılık Fakültesi Analitik Kimya Ana Bilim Dalı Adana

³İnönü Üniversitesi Eczacılık Fakültesi Farmasötik Teknoloji Ana Bilim Dalı Malatya

⁴Anadolu Üniversitesi Yunus Emre Sağlık Hizmetleri Meslek Yüksekokulu Eczacılık Bölümü Eczane Hizmetleri Programı Eskişehir

⁵Hacettepe Üniversitesi Eczacılık Fakültesi Farmasötik Teknoloji Ana Bilim Dalı Ankara

GİRİŞ ve AMAÇ: Bu çalışmanın amacı, HPLC ve UV-VIS Spektrofotometrisi kullanılarak piroksikamın asit ayrışma sabiti (pKa) ve piroksikamın ayrılma katsayısı (Log P), dağılım katsayısı (Log D) ve "Log kw" değerlerini HPLC kullanarak belirlemektir.

YÖNTEM ve GEREÇLER: HPLC çalışmaları, 1.0 mL min⁻¹ akış hızında ters faz kromatografisinde ACE C18 (150 x 4.6 mm ID, 5 µm) kolon kullanılarak gerçekleştirildi. Detektör 360 nm'ye ayarlandı. Farklı pH değerlerinde (3.0 - 6.5) log D değeri, hareketli faz olarak fosfat tamponu (20 mM): asetonitril (30: 70 v/v) karışımı ile incelendi. pKa'nın belirlenmesi için HPLC çalışmaları, 3.50 ve 6.00 pH aralığında fosfat tamponu (20 mM): metanol karışımı ile gerçekleştirildi. Log kw ölçümleri, pH 3.50 ile 6.00 arasında bir pH aralığında fosfat tamponu (20 mM): MeOH (20: 80 v/v ile 10: 90 v/v) karışımları ile yapıldı. UV-GB Spektrofotometrik pKa ölçümleri 285 nm dalga boyunda gerçekleştirildi.

BULGULAR: Piroksikamın pKa değeri sırasıyla HPLC ile 5.3 ve UV-VIS spektrofotometresi ile 5.7 olarak bulundu. Deney şartlarımızda piroksikamın log P değeri 1.58 olarak bulundu. Log D değerleri, sırasıyla 3.17, 3.79, 4.44, 5.42 ve 6.56 pH değerleri için 1.57, 1.57, 1.44, 1.13 ve 0.46 olarak bulundu.

TARTIŞMA ve SONUÇ: Literatürde, piroksikam için farklı Log P (3.1, 2.2 ve 0.6) ve pKa (6.3 ve 4.8) değerleri bildirilmiştir. Çalışmamızda piroksikam için elde edilen Log P (1.58) ve pKa (5.3 ve 5.7) değerleri literatür değerleri arasındaydı. Tüm bu sonuçlar, fizikokimyasal özelliklerin belirlenmesinde kullanılan farklı deneysel yaklaşımların farklı değerler sağlayabileceğini göstermektedir. UV Spektrofotometrisinin uygulanması kolay olsa da, HPLC, bileşiklerin pKa, Log D ve Log P değerlerinin eşzamanlı olarak belirlenmesi için eşsiz tekniklerden biridir.

Anahtar Kelimeler: Piroksikam, fizikokimyasal özellikler, pKa, Log P, Log D, Log kw

Determination of the physicochemical properties of piroxicam

Mustafa Çelebier¹, Merve Nenni², OZAN KAPLAN¹, emrah akgeyik³, Mustafa Sinan Kaynak⁴, Selma Şahin⁵

¹Hacettepe University, Faculty of Pharmacy, Department of Analytical Chemistry, Ankara, Turkey

²Cukurova University, Faculty of Pharmacy, Department of Analytical Chemistry, Adana, Turkey

³Inönü University, Faculty of Pharmacy, Department of Pharmaceutical Technology, Malatya, Turkey

⁴Anadolu University, Yunus Emre Vocational School, Department of Pharmacy, Program in Pharmacy Services, Eskişehir, Turkey

⁵Hacettepe University, Faculty of Pharmacy, Department of Pharmaceutical Technology, Ankara, Turkey

INTRODUCTION: The aim of this study was to determine the acid dissociation constant (pKa) of piroxicam using HPLC and UV-VIS Spectrophotometry, and to determine the partition coefficient (Log P), distribution coefficient (Log D) and "Log kw" values of piroxicam using HPLC method.

METHODS: The HPLC studies were performed on a reversed-phase ACE C18 (150 x 4.6 mm ID, 5 µm) column at a flow rate of 1.0 mL min⁻¹. The detector was set at 360 nm. Log D at different pH values (3.0 – 6.5) was examined with a phosphate buffer (20 mM): acetonitrile (30: 70 v/v) mixture as the mobile phase. For pKa determination, HPLC studies were performed with a mixture of phosphate buffer (20 mM): methanol within the pH range of 3.50 and 6.00. Log kw measurements were performed with phosphate buffer (20 mM): MeOH (from 20: 80 v/v to 10: 90 v/v) mixtures within the pH range of 3.50 and 6.00. UV-VIS Spectrophotometric pKa measurements were performed at 285 nm wavelength.

RESULTS: pKa value of piroxicam was found to be 5.3 by HPLC and 5.7 by UV-VIS spectrophotometry, respectively. Log P of piroxicam was determined as 1.58 in our experimental conditions. Log D values were 1.57, 1.57, 1.44, 1.13 and 0.46 for pH values of 3.17, 3.79, 4.44, 5.42 and 6.56, respectively.

DISCUSSION AND CONCLUSION: In the literature, different Log P (3.1, 2.2 and 0.6) and pKa (6.3 and

4.8) values were reported for piroxicam. The Log P (1.58) and pKa (5.3 and 5.7) values obtained for piroxicam in our study were within the range of literature values. All these results indicate that different experimental approaches used for determination of physicochemical properties could provide different values. Although UV Spectrophotometry is easy to apply, HPLC is one of the unique techniques for simultaneous determination of pKa, Log D and Log P values of compounds.

Keywords: Piroxicam, physicochemical properties, pKa, Log P, Log D, Log kw

Introduction

Dissolution from the dosage form is one of the factors that limits the absorption of drugs from the gastrointestinal (GI) tract. According to the Noyes-Whitney dissolution model (1), the *in vivo* dissolution rate is influenced by drug diffusivity, drug solubility in gastrointestinal contents, wetted surface area of solid by biological fluids, and the gastrointestinal hydrodynamics (2).

Nonsteroidal anti-inflammatory drugs (NSAIDs) are chronically used as anti-inflammatory, analgesic, and antipyretic agents (3) to reduce pain, decrease stiffness, and improve functions of patients suffering from all forms of arthritis. They are also used for the acute treatment of pain associated with headache, dysmenorrhea and postoperative pain (4). Recent studies have focused on the usage of NSAIDs for cancer treatment or prevention from cancer (5). Pharmacologic treatment of cancer pain using NSAIDs has also been investigated for a long time (6), although their use is limited to their side effects (7). The relationship between some gastrointestinal tract diseases and NSAIDs usage is also another discussion (8). Drug-drug interactions related with the usage of NSAIDs are still an issue to be investigated (9). Because of all these aspects, the physicochemical properties of drugs are one of the key points to understand their behavior inside the body. Therefore, to understand the drug's behavior in GI tract, it is important to know the lipophilicity and pKa of drugs (10).

Piroxicam belongs to the non-steroidal anti-inflammatory drug (NSAID) of the oxicam class, and is used to alleviate the symptoms of painful and inflammatory conditions such as arthritis (3). They are also used for the treatment of headache, dysmenorrhea, and postoperative pain (4).

In the literature, it is easy to find various studies for determination of the pKa values of pharmaceuticals using HPLC (11-14), UV spectrophotometry (15-17), capillary electrophoresis (18, 19) and potentiometric titrations (20-22). Identical analytical techniques have also been applied over a long period of time for calculation of Log P and Log D values (23-30). For piroxicam, the reported Log P values differ from 0.6 to 3.6, and the reported pKa values differ from 4.76 to 6.30 according to the Drugbank.

Variable results in such basic physicochemical parameters motivated us to investigate these parameters in our conditions. In this study, a simple experimental procedure based on HPLC and UV-VIS Spectrophotometry was applied for determination of physicochemical properties (pKa, Log P and Log D) of piroxicam (Figure 1). Comparison of HPLC and UV-VIS Spectrophotometry used for determination of pKa values were also carried out. A chromatographic approach based on the “Log kw” is suggested in the literature (31, 32) to determine the lipophilicity of drugs. This technique is relatively new than classical shake flask method (33), and to our knowledge, there is no report available in the literature to correlate the shake flask method and Log kw measurements with each other. So that, this is the only study to compare the Log P and Log kw measurements for an active pharmaceutical ingredient.

Materials and Methods

Chemicals

Piroxicam was supplied by Sigma Aldrich (St. Louis, MO, USA). Sodium dihydrogen phosphate ($\text{NaHPO}_4 \cdot 2\text{H}_2\text{O}$) and NaOH were purchased from Merck (Darmstadt, Germany). Disodium hydrogen phosphate ($\text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$), potassium chloride (KCl), acetonitrile (ACN), methanol (MeOH) and 1-octanol were obtained from Sigma Aldrich. Water was obtained from the Milli-Q water system (Barnstead, USA) and used for preparation of all standard solutions and buffers.

Instrumentation

The HPLC system equipped with a gradient pump (Spectra-SYSTEM P2000), a degasser (Spectra SYSTEM SCM 1000), a manual injector with 20 μL injection loop (Rheodyne), and a detector (SpectraSYSTEM UV2000, Thermo Separation Products, USA) was used. The detector was adjusted to 360 nm, and retention times were determined automatically by an online computer equipped with ChromQuest software. The separations were performed using a reversed-phase ACE C18 (125 x 4.6 mm ID, 5 μm) HPLC column (Aberden, Scotland) at flow rate of 1 mL min^{-1} . The spectrophotometric measurements were carried out using a Varian Cary 50 UV-Vis Wavelength Spectrophotometer with The Xenon lamp (200 – 800 nm). The UV spectra of reference and sample solutions were determined in 1 cm quartz cells from 200 to 800 nm wavelengths.

Solutions

Standard stock solution of Piroxicam (1000 µg mL⁻¹ in MeOH)

Piroxicam standard stock solution was prepared by dissolving piroxicam (50 mg) in MeOH in a volumetric flask (50 mL).

Phosphate buffer (20 mM) with potassium chloride (100 mM) solution (between pH 3.00 and 7.50) for UV-VIS Spectrophotometric pKa determination

Phosphate buffer (20.0 mM) and 100 mM KCl mixture were prepared by dissolving 3.56 g of sodium dihydrogen phosphate (NaH₂PO₄ • 2H₂O) and 7.45 g of potassium chloride (KCl) in about 800 mL water, and then making up the volume to 1000 mL with water. The pH of the solutions was adjusted with 1 M NaOH and 0.1 M KCl mixture solution.

Phosphate buffer (20 mM): MeOH (50:50 v/v) solutions (between pH 3.50 and 6.00) for HPLC pKa determination

Phosphate buffer (20.0 mM) was prepared by dissolving 3.12 g of sodium dihydrogen phosphate (NaH₂PO₄ • 2H₂O) in about 800 mL water, and then making up the volume to 1000 mL with water. The mobile phase was prepared as 500 mL mixture of phosphate buffer/methanol (50:50 v/v), and adjusting the pH with o-phosphoric acid/MeOH (50:50 v/v) or 20 mM disodium hydrogen phosphate (Na₂HPO₄)/MeOH (50:50 v/v) mixtures for different pH values in range of 3.64-5.94.

Phosphate buffer (20 mM) solutions (between pH 3.00 and 6.60) for Log D determination

Phosphate buffer (20.0 mM) was prepared by dissolving 3.56 g of disodium hydrogenphosphate (Na₂HPO₄ • 2H₂O) in about 800 mL water, and then making up the volume to 1000 mL with water, and adjusting the pH with o-phosphoric acid to different pH values in a range of 3.17-6.56.

Phosphate buffer (20 mM, pH 3.0): ACN (30:70 v/v) solutions as the HPLC mobile phase on Log D determination

Phosphate buffer (20.0 mM) was prepared by dissolving 3.56 g of disodium hydrogenphosphate (Na₂HPO₄ • 2H₂O) in about 800 mL water, and adjusting the pH with o-phosphoric acid to 3.0, and then making up the volume to 1000 mL with water. The mobile phases were prepared as 500 mL mixture of phosphate buffer (20 mM, pH 3.0): ACN (30:70 v/v).

Phosphate buffer (20 mM): MeOH (from 20:80 v/v to 10:90 v/v) solutions for Log kw determination (between pH 3.00 and 6.00)

Phosphate buffer (20.0 mM) was prepared by dissolving 3.56 g of disodium hydrogenphosphate ($\text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$) in about 800 mL water, and then making up the volume to 1000 mL with water. The mobile phases were prepared as 500 mL mixture of phosphate buffer (20 mM)/MeOH in a range of 20:80-10:90, v/v. The pH of the mobile phases were adjusted with o-phosphoric acid/methanol (20:80 – 10:90, v/v) or 20 mM disodium hydrogen phosphate (Na_2HPO_4)/methanol (20:80 – 10:90, v/v) mixtures for different pH values (3.00 and 6.00) and different mobile phase compositions (phosphate buffer (20 mM)/MeOH in a range of 20:80 – 10:90, v/v).

Procedures

UV-VIS Spectrophotometric pKa determination

Standard stock solution of piroxicam was diluted to $20.0 \mu\text{g mL}^{-1}$ by using Phosphate buffer (20 mM) potassium chloride (100 mM) solutions for UV-VIS Spectrophotometric pKa determination (pH 3.09 – 7.48). Spectra were taken from 200 to 400 nm wavelength. The shifts on absorption were considered. A sigmoidal curve was constructed between pH of the solutions and absorption at 285 nm. The pKa of piroxicam was determined according to the sigmoidal relationship given by Microsoft Excel between pH 3.09 and 7.48.

HPLC pKa determination

Standard stock solution of piroxicam was diluted to $5.0 \mu\text{g mL}^{-1}$ with phosphate buffer (20 mM): MeOH (50:50 v/v) solutions for HPLC pKa determination (pH 3.64 – 5.94). These solutions were injected into the HPLC system with the mobile phases in which they were dissolved. A sigmoidal curve was constructed between the pH of the mobile phases and capacity factor (k') of piroxicam. The pKa of piroxicam was determined according to the sigmoidal relationship between pH 3.64 and 5.94.

Log D determination

The experimental system was a modified shake-flask method to meet the requirements of OECD guideline for testing of chemicals (33). For this purpose, standard stock solution of piroxicam was diluted to $250 \mu\text{g mL}^{-1}$ with MeOH. $50.0 \mu\text{L}$ of this solution was added into aqueous biphasic systems containing phosphate buffer (20 mM)

solutions for Log D determination (pH 3.17 – 6.56) and 1-octanol (50:50 v/v, 950 μL). After the extraction at ambient temperature, the amount of piroxicam remaining in buffer was determined by HPLC. Phosphate buffer (20 mM, pH 3.0): ACN (30:70 v/v) mixture was used as mobile phase and the buffer phase of the extracted sample was diluted 5-fold with the mobile phase before injection into the HPLC system. Peak areas were compared with 5.0 $\mu\text{g mL}^{-1}$ piroxicam solution to determine the piroxicam amount that partition into the octanol phase.

Log P determination

The lipophilicity of a compound can be expressed by its partition coefficient (Log P) which is the concentration ratio of a non-ionized compound in a mixture of two immiscible phases (aqueous and organic phases) at equilibrium. For measurement of the Log P value of a weak basic drug, piroxicam, the equation given below is used for different pH values. The average of the obtained results were calculated to find the Log P value.

$$\text{Log } D = \text{Log } P - \text{Log}(1 + 10^{pK_a - pH}) \quad \text{Equation 1}$$

Log k_w determination

For determination of Log k_w values, piroxicam was diluted to 5.0 $\mu\text{g mL}^{-1}$ using phosphate buffer (20 mM): MeOH (from 20:80 v/v to 10:90 v/v) solutions (pH 3.00 and 6.00). The relationship between Log k' and methanol concentration in the mobile phase is described with $\text{Log } k' = \text{Log } k_w - S\phi$ (31, 34). In this equation, k_w is the k' value for a compound when aqueous phase is used as the eluent, S is the slope of the regression line, ϕ is the volume percentage of methanol in the mobile phase. If ϕ is zero (no methanol in the mobile phase), the mobile phase is consisted of only the phosphate buffer, the Log k' will be equal to the Log k_w .

Measurements on Sigmoidal Curves

The data measured from sigmoidal curve for pKa determination were found using two different approaches. The first one was to print the whole graph in a A4 paper and finding the pKa values by drawing tangents. The second one was to use the derivative of the actual graph.

Results and Discussion

In this study, we focused on the pKa, Log P and Log D values of piroxicam because the reported values are very different from each other. Log kw values of piroxicam were also evaluated and the values were compared with the Log D values.

UV-VIS Spectrophotometric pKa determination

Absorption of organic compounds including most of the drugs is based on transitions of n or π electrons to the π^* excited state. The absorption peaks for these transitions fall in 200 - 700 nm range which is experimentally convenient region of the spectrum. Referred transitions require an unsaturated group in the molecule to provide the π electrons. The solvent in which the absorbing species are dissolved may have serious effects on the spectrum (35). In the present study, the spectra of piroxicam in the phosphate buffers having identical ionic strengths (adjusted with 0.1 M KCl) but at different pH values from 3.09 to 7.48 were recorded. The values between 3.09 and 7.48 were used to construct the sigmoidal curve at 285 nm wavelength. The characteristic of absorption spectrum for piroxicam was changed by changing the pH of the buffered aqueous media (Figure 2). The changes in the absorbance values are usually followed by overlaid plots of recorded spectra, and the greatest change occurs when the acidity of the aqueous solution is equal to the pKa of the studied compound. The sigmoidal curve was constructed and pKa value of piroxicam was estimated as 5.7 (Figure 3.)

HPLC pKa determination

The capacity factor of a drug in a reversed phase HPLC system is related with its lipophilicity (36, 37). Since weak acidic drugs like piroxicam are ionized at basic pH, they tend to be eluted rapidly from a lipophilic C₁₈ stationary phase with basic mobile phases. This situation is vice versa for acidic mobile phases. Our results are in a good agreement with this statement. Piroxicam was eluted at 7.44 min at acidic pH (pH 3.64) and eluted at 4.34 min at relatively basic pH (pH 5.94) values (Figure 4). When the pH value of mobile phase was plotted against k' values, a sigmoidal relationship was obtained between pH and capacity factor (Figure 5). The pK_a value of piroxicam was calculated as 5.3 from this relationship.

Log D and Log P determination

In drug discovery and development, lipophilicity of a compound is usually expressed by its partition between water and 1-octanol. The concentration of a non-ionized

compound in the organic and aqueous phases refers to Log P. The log P of any ionizable solution can be measured in the aqueous phase in which the pH is adjusted to the non-ionized form. The concentration ratio of non-ionized solute in the solvents is calculated according to the Log P value which is a measure of lipophilicity, and is not pH dependent (Equation 2).

$$\text{Log } P_{\text{oct/wat}} = \text{Log} \left(\frac{[\text{solute}]_{\text{octanol}}}{[\text{solute}]_{\text{un-ionized water}}} \right) \quad \text{Equation 2}$$

As described above, piroxicam is a weak base, and it is partially ionized when dissolved in water. Log D is the ratio of the sum of the concentrations of all forms of the compound (ionized plus non-ionized) in each phase, and is pH dependent. The distribution coefficient is defined as a function of the ratio of total concentration of the solute species in each phase (Equation 3) (38).

$$\text{Log } D_{\text{oct/wat}} = \text{Log} \left(\frac{[\text{solute}]_{\text{octanol}}}{[\text{solute}]_{\text{ionized water}} + [\text{solute}]_{\text{neutral water}}} \right) \quad \text{Equation 3}$$

For a non-ionized drug, Log P is equal to Log D at any pH value, but the effective lipophilicity at any specific pH value is directly related to its pKa value for any ionized drug. In our experimental conditions, Log D of piroxicam were investigated at pH 3.17, 3.79, 4.44, 5.42 and 6.56. Table 1 summarizes the Log D values of piroxicam at the investigated pH values. Log P values given in Table 1 were calculated according to the Equation 1 where pKa of the piroxicam was accepted as 5.7 and 5.3 by the results of UV Spectrophotometric and HPLC measurements, respectively.

Since piroxicam is an acidic drug, it is ionized at basic pH values and tends to be dissolved in aqueous phases. As it was expected, solubility of piroxicam in aqueous phase (phosphate buffer) was increased about 13 fold by the changes at pH from 6.56 to 3.17. This situation was in a harmony with the capacity factor of piroxicam on HPLC for different mobile phase pH values. Although Log D values differ by pH values, calculated Log P values should be identical according to the theory. When the mean and standard error of Log P values were calculated it was found that Log P was 1.58 ± 0.04 if pKa is taken as 5.3, and 1.46 ± 0.05 if pKa is taken as 5.7. This situation shows us that the pKa value of 5.3 determined by HPLC, provides less standard error for the

calculated Log P values. Therefore, Log P value for piroxicam was accepted as 1.58 in our experimental conditions.

Log kw determination

Since the retention time of an analyte in RP-HPLC is directly related with its lipophilicity, this relationship can be used to show how the lipophilicity of an analyte will be affected by pH changes. In our study, initially we have calculated the Log k' values for all mobile phase compositions (80:20, 85:15, 90:10 MeOH:phosphate buffer (20 mM); (v/v)) for pH values below (3.0) and above (6.0) the calculated pKa (5.3). The Log k' values were calculated for the mobile phase not containing any organic phase (0 % MeOH). Figure 5 shows an example of this application for pH 6.0. When there was no MeOH in the mobile phase, the k_w' values were calculated for pH 3.0 and 6.0 and found to be 37.5 and 7247.7, respectively.

Conclusion

In this study, spectrophotometric and chromatographic analytical approaches were examined to determine the physicochemical properties of piroxicam. Since, piroxicam is a well-known compound, the results were easily compared with the ones in Drugbank (<https://www.drugbank.ca/drugs/DB00554>). According to our experimental results, pKa value was found to be 5.7 and 5.3 for spectrophotometric and HPLC experiments, respectively. The value found by UV-VIS Spectrophotometry is close to experimental value of 6.3 given in Drugbank. However, the predicted value of 4.76 given in Drugbank is close to the one calculated by the results of HPLC analysis. Experimental Log P value in Drugbank for piroxicam is 3.06 where predicted values are 0.6 and 2.2. Our experiments show that piroxicam Log P value is 1.58. This value is between the predicted and experimental values reported in Drugbank. The differences for pKa and Log P between experimental and predicted values in Drugbank, indicate the experimental data found in this study is novel for piroxicam. Based on all these results, we can conclude that use of different experimental approaches for determination of physicochemical properties can clearly provide different values for drugs.

REFERENCES

1. Costa P, Lobo JMS. Modeling and comparison of dissolution profiles. European journal of pharmaceutical sciences. 2001;13(2):123-33.

2. Hörter D, Dressman J. Influence of physicochemical properties on dissolution of drugs in the gastrointestinal tract. *Advanced drug delivery reviews*. 2001;46(1):75-87.
3. Crofford LJ. Use of NSAIDs in treating patients with arthritis. *Arthritis Res Ther*. 2013;15(Suppl 3):S2.
4. Simon LS. Nonsteroidal anti-inflammatory drugs and their risk: a story still in development. *Arthritis Res Ther*. 2013;15(Suppl 3):S1.
5. Cha YI, DuBois RN. NSAIDs and cancer prevention: targets downstream of COX-2. *Annu Rev Med*. 2007;58:239-52.
6. Portenoy RK, Lesage P. Management of cancer pain. *The Lancet*. 1999;353(9165):1695-700.
7. Carr DB, Goudas LC, Balk EM, Bloch R, Ioannidis J, Lau J. Evidence report on the treatment of pain in cancer patients. *Journal of the National Cancer Institute Monographs*. 2003(32):23-31.
8. Rainsford KD, Velo GP. *Side-Effects of Anti-Inflammatory Drugs: Part Two Studies in Major Organ Systems*: Springer Science & Business Media; 2012.
9. Franz CC, Egger S, Born C, Bravo AER, Krähenbühl S. Potential drug-drug interactions and adverse drug reactions in patients with liver cirrhosis. *European journal of clinical pharmacology*. 2012;68(2):179-88.
10. Dressman J, Vertzoni M, Goumas K, Reppas C. Estimating drug solubility in the gastrointestinal tract. *Advanced drug delivery reviews*. 2007;59(7):591-602.
11. Uhrova M, Miksik I, Deyl Z, Bellini S. Determination of dissociation constants by separation methods (HPLC and CE). Theoretical background and guidelines for application. *Process Contr Qual*. 1997;10(1-2):151-67.
12. Oumada FZ, Rafols C, Roses M, Bosch E. Chromatographic determination of aqueous dissociation constants of some water-insoluble nonsteroidal antiinflammatory drugs. *J Pharm Sci*. 2002;91(4):991-9.
13. Demiralay EC, Koc D, Daldal YD, Cakir C. Determination of chromatographic and spectrophotometric dissociation constants of some beta lactam antibiotics. *J Pharmaceut Biomed*. 2012;71:139-43.
14. Canbay HS, Demiralay EC, Alsancak G, Ozkan SA. Chromatographic Determination of pK(a) Values of Some Water-Insoluble Arylpropionic Acids and Arylacetic Acids in Acetonitrile plus Water Media. *J Chem Eng Data*. 2011;56(5):2071-6.
15. Santos E, Rosillo I, Delcastillo B, Avendano C. Determination of Pka Values for Hydantoins by Spectrophotometry. *J Chem Res-S*. 1982(5):131-.
16. Rosenberg LS, Simons J, Schulman SG. Determination of Pka Values of N-Heterocyclic Bases by Fluorescence Spectrophotometry. *Talanta*. 1979;26(9):867-71.
17. Pereira AV, Garabeli AA, Schunemann GD, Borck PC. Determination of Dissociation Constant (K-a) of Captopril and Nimesulide - Analytical Chemistry Experiments for Undergraduate Pharmacy. *Quim Nova*. 2011;34(9):1656-60.
18. Fu XF, Liu Y, Li W, Bai Y, Liao YP, Liu HW. Determination of dissociation constants of aristolochic acid I and II by capillary electrophoresis with carboxymethyl chitosan-coated capillary. *Talanta*. 2011;85(1):813-5.
19. Ehala S, Misek J, Stara IG, Stary I, Kasicka V. Determination of acid-base dissociation constants of azahelicenes by capillary zone electrophoresis. *J Sep Sci*. 2008;31(14):2686-93.

20. Schurman.H, Thun H, Verbeek F. Potentiometric Determination of Dissociation Constants of Itaconic Acid. *J Electroanal Chem.* 1970;26(2-3):299-&.
21. Qiang ZM, Adams C. Potentiometric determination of acid dissociation constants ($pK(a)$) for human and veterinary antibiotics. *Water Res.* 2004;38(12):2874-90.
22. Roda G, Dallanoce C, Grazioso G, Liberti V, De Amici M. Determination of Acid Dissociation Constants of Compounds Active at Neuronal Nicotinic Acetylcholine Receptors by Means of Electrophoretic and Potentiometric Techniques. *Anal Sci.* 2010;26(1):51-4.
23. Mirrlees MS, Moulton SJ, Murphy CT, Taylor PJ. Direct measurement of octanol-water partition coefficients by high-pressure liquid chromatography. *Journal of medicinal chemistry.* 1976;19(5):615-9.
24. Haky JE, Young AM. Evaluation of a simple HPLC correlation method for the estimation of the octanol-water partition coefficients of organic compounds. *Journal of liquid chromatography.* 1984;7(4):675-89.
25. Kaliszan R, Haber P, Baczek T, Siluk D. Gradient HPLC in the determination of drug lipophilicity and acidity. *Pure and Applied Chemistry.* 2001;73(9):1465-75.
26. Yamana T, Tsuji A, Miyamoto E, Kubo O. Novel method for determination of partition coefficients of penicillins and cephalosporins by high - pressure liquid chromatography. *Journal of pharmaceutical sciences.* 1977;66(5):747-9.
27. Wiczling P, Kawczak P, Nasal A, Kaliszan R. Simultaneous determination of pK_a and lipophilicity by gradient RP HPLC. *Analytical chemistry.* 2006;78(1):239-49.
28. Singh S, Sharda N, Mahajan L. Spectrophotometric determination of pK_a of nimesulide. *International journal of pharmaceutics.* 1999;176(2):261-4.
29. Völgyi G, Ruiz R, Box K, Comer J, Bosch E, Takács-Novák K. Potentiometric and spectrophotometric pK_a determination of water-insoluble compounds: Validation study in a new cosolvent system. *Analytica chimica acta.* 2007;583(2):418-28.
30. Cleveland Jr J, Benko M, Gluck S, Walbroehl Y. Automated pK_a determination at low solute concentrations by capillary electrophoresis. *Journal of Chromatography A.* 1993;652(2):301-8.
31. Hong H, Wang L, Zou G. Retention in RP-HPLC: Lipophilicity determination of substituted biphenyls by reversed-phase high performance liquid chromatography. *Journal of liquid chromatography & related technologies.* 1997;20(18):3029-37.
32. Markuszewski MJ, Wiczling P, Kaliszan R. High-throughput evaluation of lipophilicity and acidity by new gradient HPLC methods. *Combinatorial chemistry & high throughput screening.* 2004;7(4):281-9.
33. Kocak E, Celebier M, Altinoz S. Validation of spectrophotometric method to quantify varenicline content in tablets. *Asian Journal of Chemistry.* 2013;25(4):1845-8.
34. Braumann T. Determination of hydrophobic parameters by reversed-phase liquid chromatography: theory, experimental techniques, and application in studies on quantitative structure-activity relationships. *Journal of Chromatography A.* 1986;373:191-225.

35. Celik H, Buyukaga M, Celebier M, Turkoz Acar E, Baymak MS, Gokhan-Kelekci N, et al. Determination of pKa values of some benzoxazoline derivatives and the structure-activity relationship. *Journal of Chemical and Engineering Data*. 2013;58(6):1589-96.
36. Demiralay EC, Alsancak G, Ozkan SA. Determination of pKa values of nonsteroidal antiinflammatory drug-oxicams by RP-HPLC and their analysis in pharmaceutical dosage forms. *Journal of separation science*. 2009;32(17):2928-36.
37. Wiczling P, Kawczak P, Nasal A, Kaliszan R. Simultaneous determination of pKa and lipophilicity by gradient RP HPLC. *Anal Chem*. 2006;78(1):239-49.
38. Royal Society of Chemistry, Learn Chemistry, Partition and distribution coefficients <http://www.rsc.org/learn-chemistry/resource/> [

Table 1. Experimental Log D and calculated Log P values of piroxicam

pH	D	Log D	Log P (pKa is accepted as 5.3)	Log P (pKa is accepted as 5.7)
3.17	37.17	1.57	1.57	1.57
3.79	37.38	1.57	1.58	1.58

4.44	27.65	1.44	1.50	1.46
5.42	13.58	1.13	1.50	1.32
6.56	2.89	0.46	1.74	1.38

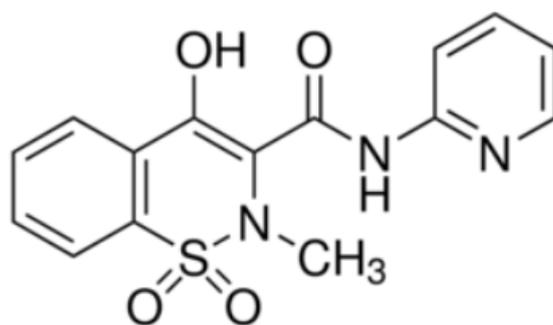


Figure 1. Chemical structure of piroxicam

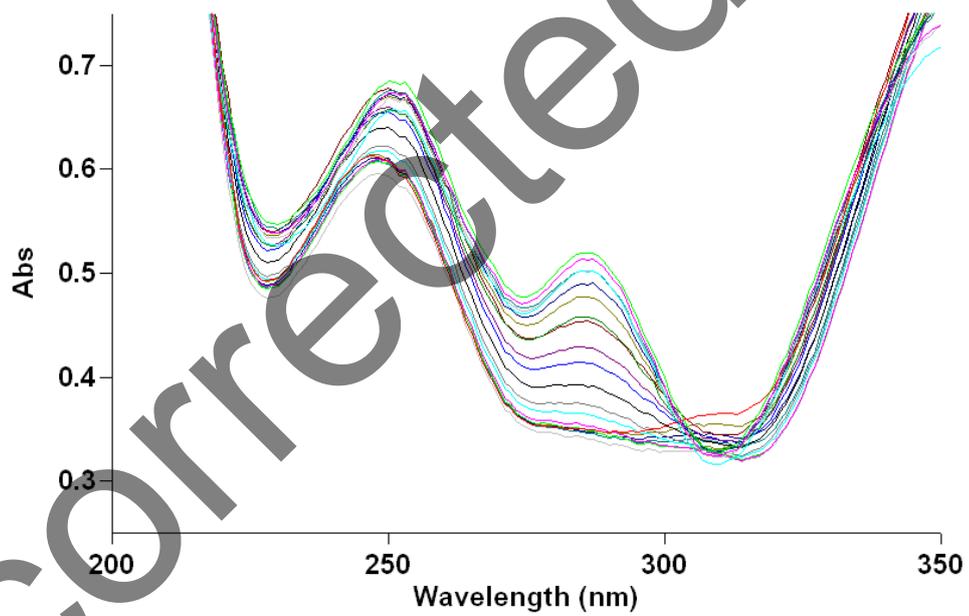


Figure 2. Representative overlaid spectra (200 – 400 nm) of piroxicam under optimum conditions at various pH values (pH 3.09 – 7.48)

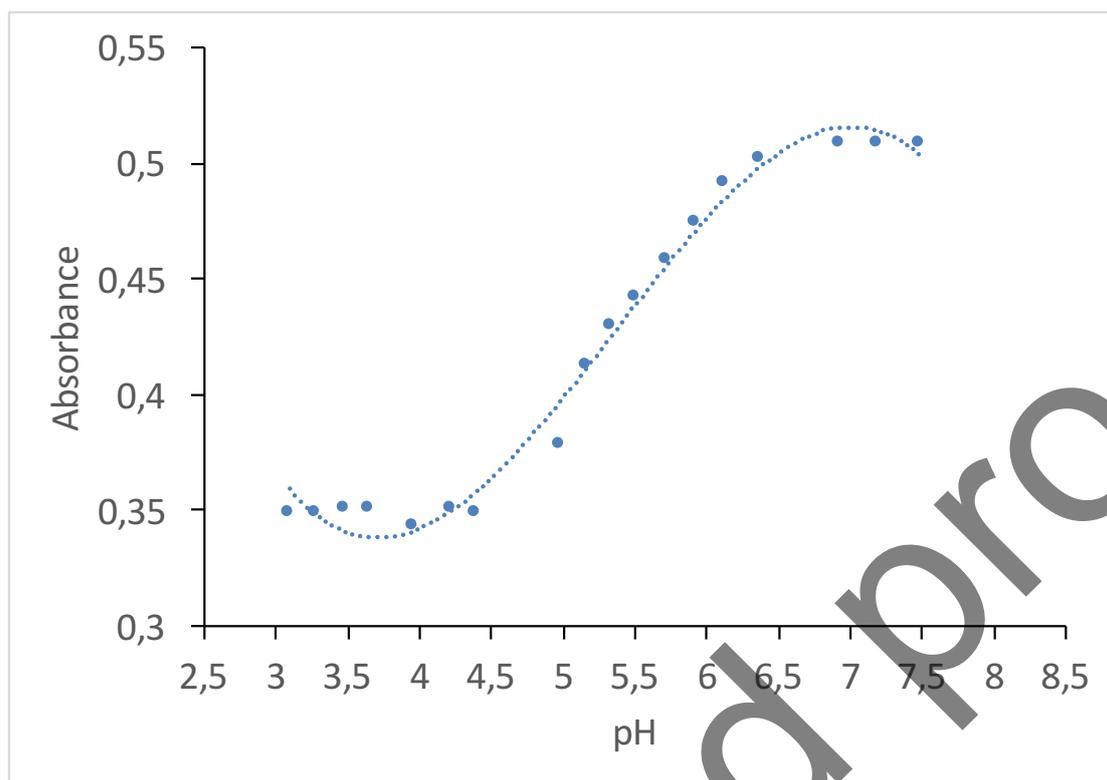


Figure 3. The plot of the absorbance values of piroxicam at 285 nm obtained as a function of pH (3.09 – 7.48)

Uncorrected proof

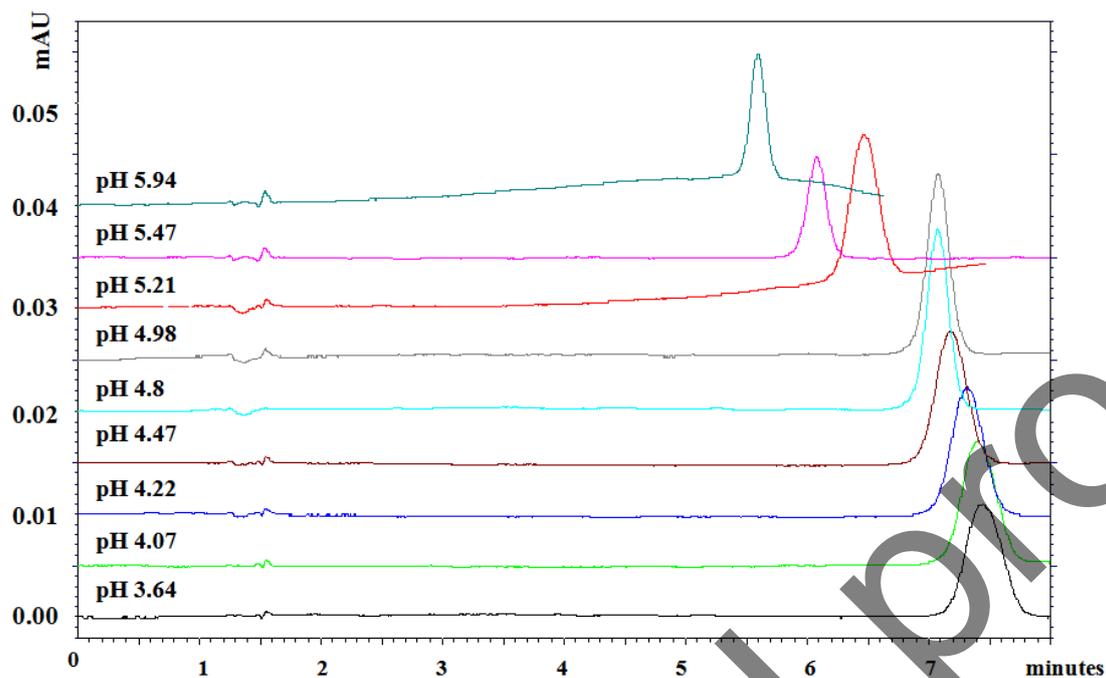


Figure 4. Representative overlaid chromatograms of piroxicam taken under optimum conditions at various pH (3.64 – 5.94). pH values were given on the top of each chromatogram peak.

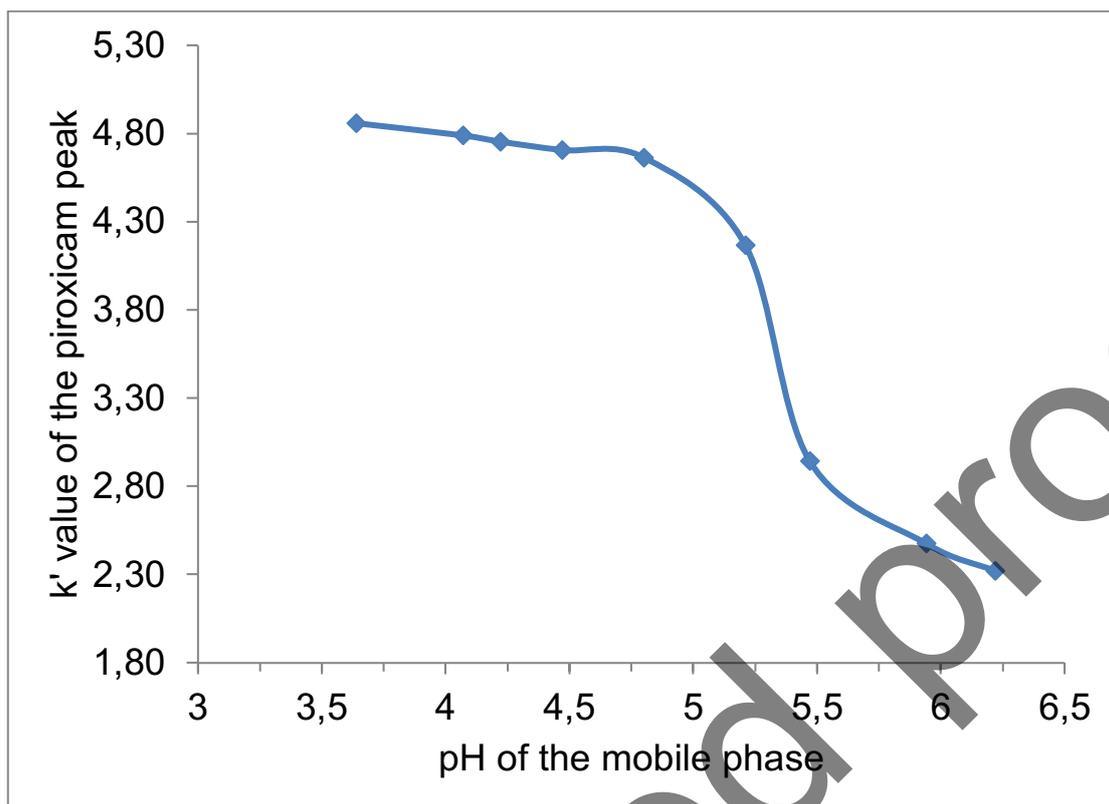


Figure 5. Sigmoidal relationship between pH (3.64 – 6.22) and k' of the piroxicam peak